

WRLFMD Quarterly Report October to December 2018

Foot-and-Mouth Disease











CONTENTS

1. Sumi	mary of samples tested and reported FMD outbreaks	. 3
1.1. As	sia	. 3
1.2. Af	rica	. 4
1.3. Sc	outh America	. 7
1.4. Ur	ncharacterised FMD viruses	. 7
2. Deta	iled Analysis	. 9
2.1. AS	SIA	10
2.2. AF	FRICA	15
2.3. Va	accine matching2	21
Annex 1: S	Sample data2	22
2.4. Su	ummary of Submissions2	22
2.5. Cl	linical Samples2	23
2.6. Ar	ntigenic Characterisation2	26
Annex 2: F	FMD publications2	29
Annex 3: V	/accine Recommendations	35
Annex 4: C	Other WRLFMD Activities	36



1. Summary of samples tested and reported FMD outbreaks

1.1. Asia

China, People's Republic of

Two further outbreaks of **FMD type O** were reported in cattle in the Inner Mongolia and Xinjiang Autonomous Regions on the 12th and 25th October, respectively. No genotyping results have been reported.

Israel

Between 25/09/2018 and 26/12/2018, 17 outbreaks of **FMD type O** were reported in the Northern District (Hazafon). These mainly occurred in cattle, but also included wild boar and mountain gazelle (Gazella gazella). No genotyping results have been reported.

Laos

One sample was received on 14/11/2018. It was collected from cattle on 22/01/2018 (location not specified). Conventional virus isolation on primary BTy cells failed to grow any virus, but rRT-PCR was positive. Chemical transfection of LFBK cells with the RNA resulted in virus growth. VP1 sequencing of amplicons from both the RNA extracted from the original sample and from the LFBK cell culture fluid showed the virus to be **FMD type O** and genotyping revealed it to belong to the PanAsia lineage of the ME-SA topotype (see below). The two sequences were identical.

Mongolia

Three retrospective outbreaks of **FMD type O** were reported in (Bactrian?) camels (Dundgovi, 03/01/2018) and cattle (Uvurkhangai, 15/05/2018 and Arkhangai, 28/05/2018). Genotyping of O/MOG/2/2018 (camel, 03/01/2018) was reported in the previous quarterly report as ME-SA/Ind-2001e. No genotyping results have been reported on the other two outbreaks.



Thailand

Nineteen samples were received on 14/11/2018. They were collected between February 2017 and August 2018 from cattle in various regions throughout the country. Eight **FMD type O** and eight **FMD type A** viruses were isolated and FMDV genome detected in the remaining three samples by rRT-PCR. Genotyping revealed that the type O viruses belonged to the ME-SA topotype, five being the PanAsia lineage and three the Ind-2001 lineage (sublineage e) (see below). All eight of the type A viruses belonged to the ASIA topotype, Sea-97 lineage (see below).

Turkey

Four **FMD** type **O** sequences were received from the FMDI-Ankara on 18/10/2018. They were collected during 2018 from Ankara and Van provinces. Genotyping revealed them to belong to the ME-SA topotype, PanAsia-2 lineage, QOM-15 sublineage (see below).

Vietnam

A single **FMD type O** VP1 sequence was sent from Regional Animal Health Office No.6 (RAHO6) on 17/12/2018. The sample had been collected from a pig during November 2018 (the location was not specified). Genotyping showed it to belong to the SEA topotype, Mya-98 lineage (see below).

1.2. Africa

Algeria

Between 11/07/2018 and 12/11/2018, 45 outbreaks of **FMD type O** were reported in cattle, sheep and goats across northern Algeria. Previous analyses showed the viruses to belong to the EA-3 topotype (see previous quarterly report).

Burkina Faso

Eighteen samples were received on 06/12/2018. They were collected from cattle sampled between June and August 2018 in four regions (Centre, Centre-Ouest, Plateau-Central and Boucle du Mouhoun). **FMD type O** viruses were isolated from seven samples, five were FMDV-GD and the remaining six were NVD. Genotyping of the virus isolates showed the topotype to be EA-3 (see below).



Gambia

Two samples were received on the 25 October 2018. They were collected on 23/07/2018 and may have been from the reported outbreaks in Njallal Samba and Ndowen villages (Niani district, Central River Division). **FMD type O** was isolated from both samples and genotyping revealed the topotype to be EA-3 (see below).

Guinea

Between 27/07/2018 and 16/09/2018, eight outbreaks of **FMD type O** were reported in cattle in five of the seven administrative regions. Previous analyses showed the viruses to belong to the EA-3 topotype (see previous quarterly report).

Malawi

In October and November 2018, two outbreaks of **suspected FMD** were reported in Chitipa (Northern Region), however, no samples were collected. Subsequently another 42 cases were reported and samples taken on the 21st December were submitted to the SSARRL (BVI); results are awaited.

Mauritania

Seven VP1 sequences from **FMD type O** viruses were received from ANSES (European Union Reference Laboratory, FAO Reference Centre & OIE Reference Laboratory for FMD). They were collected in July 2018 from Hodh Ech Chargui region. Genotyping showed them to belong to the EA-3 topotype (see below).

Senegal

Eleven samples were received on 25/10/2018. They were collected from cattle and a pig sampled between May and July 2018 in seven different regions (Dakar, Kaolack, Kedougou, Kolda, Tambacounda, Thies and Ziguinchor). **FMD type O** was isolated from six samples and FMDV genome detected in a further three; no virus or genome was detected in the remaining two samples. Genotyping revealed the topotype to be EA-3 (see below).

Sierra Leone

Thirty four samples were received on 16/10/2018. They were collected from cattle on 28/08/2018 in the Northern and Eastern provinces. Virus could not be detected in 31 samples and in the remaining 3 cases FMDV genome was detected by rRT-PCR. Subsequently RNA from one of these (from Kono, Eastern province) was chemically



transfected into LKBK cells and a virus recovered. This was identified as **FMD type O** and genotyped as the EA-3 topotype (see below).

South Africa

Three further outbreaks due to **FMD type SAT 2** were reported in cattle in the Limpopo province. No genotyping results have been reported. Additionally, sub-clinical infection of African buffalo (Syncerus caffer) at Maruleng (Limpopo province) was reported on 12/10/2018.

South Sudan

During the previous reporting period, on 29/08/2018, a batch of 30 samples was received. They were collected from cattle between April and June 2017. Diagnostic assays failed to isolate any FMD viruses, however, FMDV genome was detected in seven samples. Using a new lineage-specific real-time RT-PCR developed by WRLFMD and NAHDIC (Ethiopia) it was shown that the genome-positive samples probably contained FMDV O/EA-3 RNA. Recently, RNA extracted from one of the FMDV-GD samples was chemically transfected into LFBK cells and a virus was recovered. The virus, from cattle sampled on the 18/05/2017 in Jonglei State, was identified as **FMD type 0** and genotyping showed the topotype to be EA-3 (see below).

Tunisia

Seven outbreaks of **FMD type O** were reported in cattle and sheep in December 2018. No genotyping results have been reported.

Zambia

Three samples were received on 14/12/2018. They were collected from cattle on 24/10/2018 (unknown location). **FMD type A** viruses were isolated from all three samples. Genotyping is in progress.

Zimbabwe

Nine outbreaks of **FMD type SAT 1** have been reported in cattle between October and December 2018 in Masvingo province. A single outbreak due to **FMD type SAT 2** was reported in Mashonaland Central in November 2018 and another untyped outbreak of FMD was reported in cattle in Mashonaland West. No genotyping results have been reported.



Comment:

Serotype O in West Africa and the Maghreb

New field outbreaks due to the O/EA-3 topotype have been widely reported in a number of West African countries. EA-3 normally occurs in East Africa (Sudan, Ethiopia and Eritrea) and spread to Egypt in 2012 (where it has persisted), Libya in 2012 (only one report), Palestine (Gaza & the West Bank; 2017) and Israel (2017). O/EA-3 was first detected in West Africa in Nigeria in 2007 and subsequently in 2009, 2011, 2014 and 2016. Outbreaks due to O/EA-3 have now been confirmed in the Cameroon (2010, 2012-2013, 2015-2016), Burkina Faso (2018), the Gambia (2018), Guinea (2018), Senegal (2018) and Sierra Leone (2018). This viral topotype has recently spread to the Maghreb (confirmed in Algeria and Mauritania in 2018 and is suspected in Tunisia and Morocco).

The indigenous West African (O/WA) topotype was also present in Nigeria during 2011 to 2014 and 2016 and it will interesting to see if both of these serotype O topotypes will continue to coexist in future years.

1.3. South America

Colombia

Between 10/08/2018 and 06/10/2018, five outbreaks of **FMD type O** were reported in cattle and pigs in the Cesar Department. On 13th September and 9th November, a further two outbreaks of **FMD type O** were reported in cattle and pigs in the La Guajira Department. No genotyping results have been reported.

1.4. Uncharacterised FMD viruses

A number of outbreaks have occurred where samples have not been sent to the WRLFMD. It is probable that the countries involved have performed their own genetic characterisation; however, through the OIE/FAO Laboratory Network we would also like to encourage the submission of samples (or complete VP1 sequences) to the WRLFMD.

An up-to-date list and reports of FMD viruses characterised by sequencing can be found at the following website: http://www.wrlfmd.org/country-reports/country-reports-2018.



Results from samples or sequences received at WRLFMD (status of samples being tested) are shown in Table 1 and a complete list of clinical sample diagnostics made by the WRLFMD from October to December 2018 is shown in Annex 1 (Summary of Submissions). A record of all samples received by WRLFMD is shown in Annex 1 (clinical samples).

Table 1: Status of sequencing of samples or sequences received by the WRLFMD from October to December 2018 (* indicates a batch carried over from the previous quarter).

WRLFMD Batch No.	Date received	Country	Serotype	No. of samples	No. of sequences	Sequencing status
WRLFMD/2018/00026	25/10/2018	Gambia	0	2	2	completed
WRLFMD/2018/00027	16/10/2018	Sierra Leone	0	1	1	completed
WRLFMD/2018/00028	25/10/2018	Senegal	0	6	6	completed
WRLFMD/2018/00029	14/11/2018	Laos	0	1	1	completed
WRLFMD/2018/00030	14/11/2018	Thailand	Ο	8	8	completed
WRLFMD/2018/00030	14/11/2018	Thailand	Α	8	8	completed
WRLFMD/2018/00031	06/12/2018	Burkina Faso	0	7	7	completed
WRLFMD/2018/00032	14/12/2018	Zambia	Α	3	-	Pending
			Total	36	33	

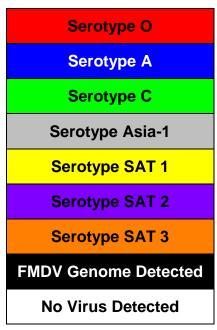
Table 2: VP1 sequences submitted by other FMD Reference laboratories to the WRLFMD from October to December 2018.

WRLFMD Batch No.	Date received	Country	Serotype	No. of sequences	Submitting laboratory
WRLMEG/2018/00037	18/10/2018	Turkey	0	4	FMDI- Ankara
WRLMEG/2018/00039	02/11/2018	Mauritania	0	7	ANSES
WRLMEG/2018/00045	17/12/2018	Vietnam	0	1	RAHO6
			Total	12	



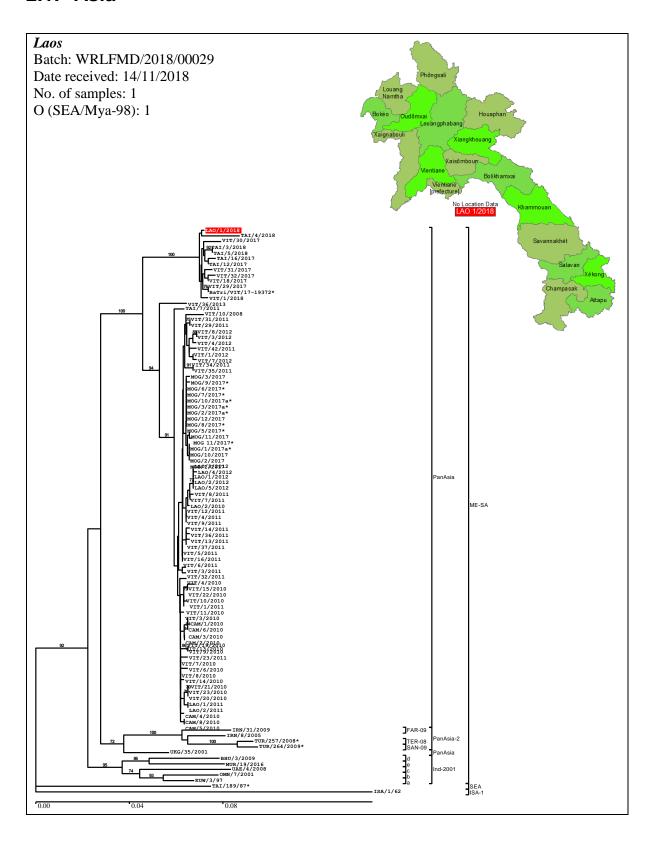
2. Detailed Analysis

Key for maps and trees:

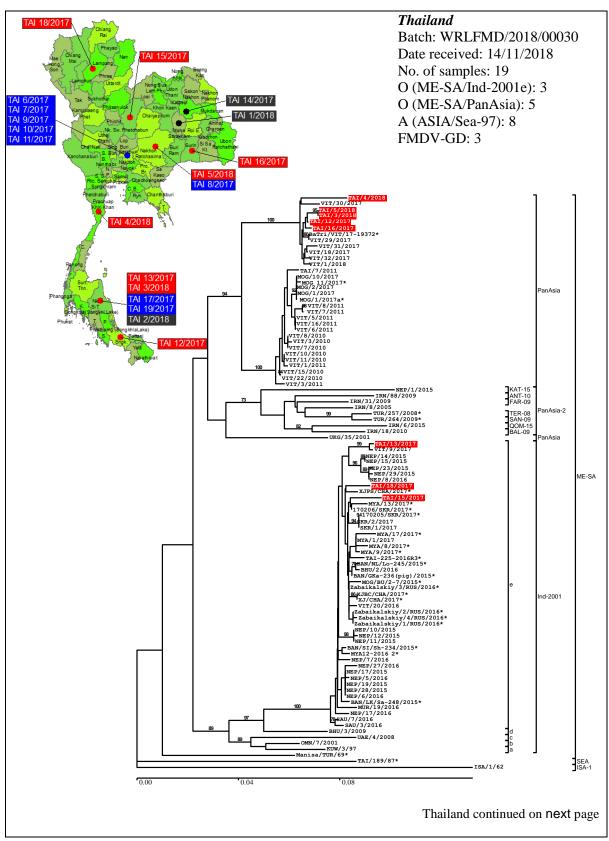




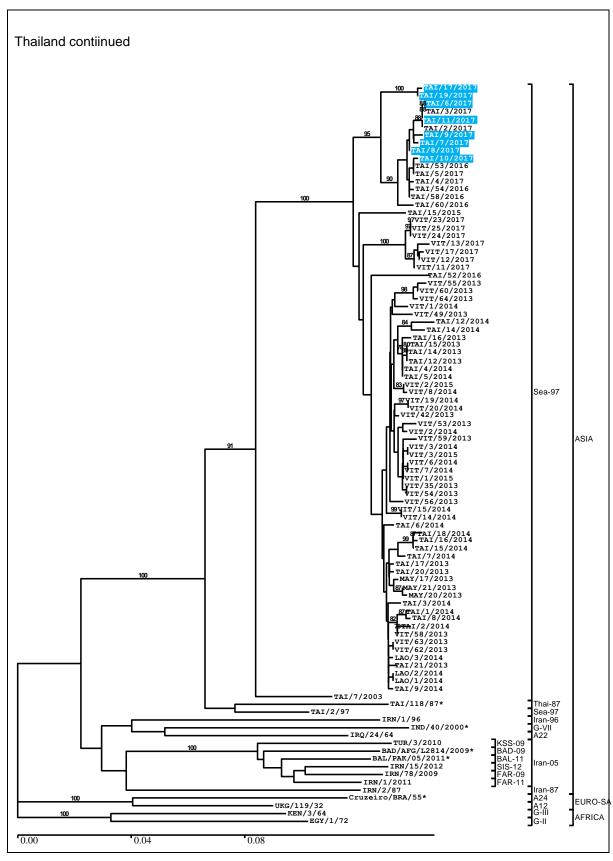
2.1. Asia



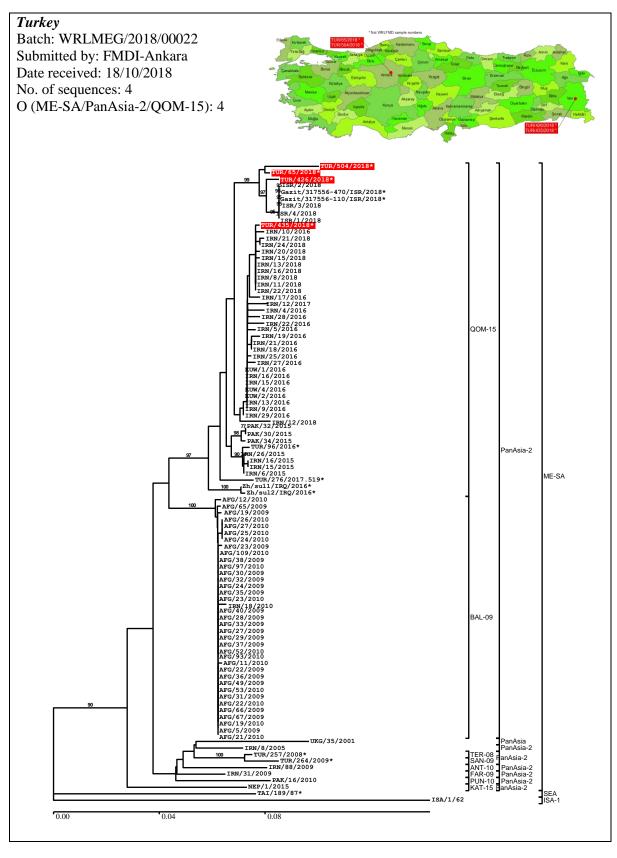




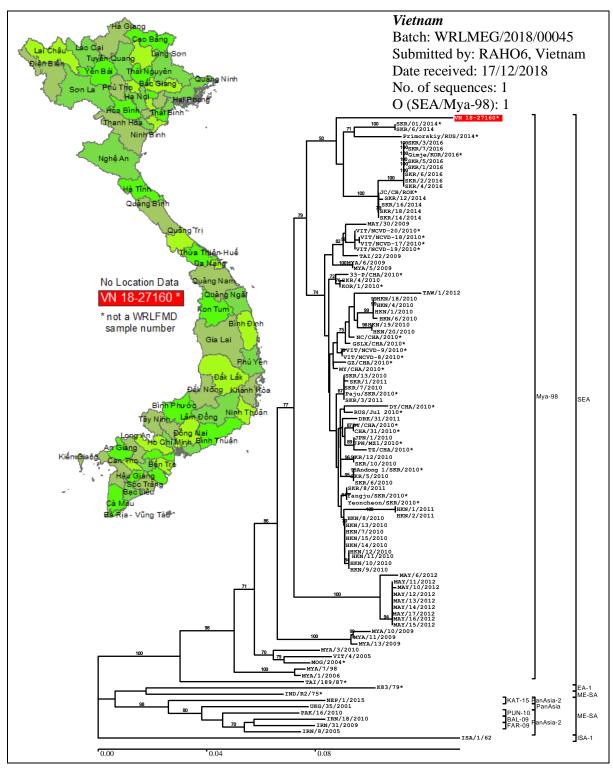






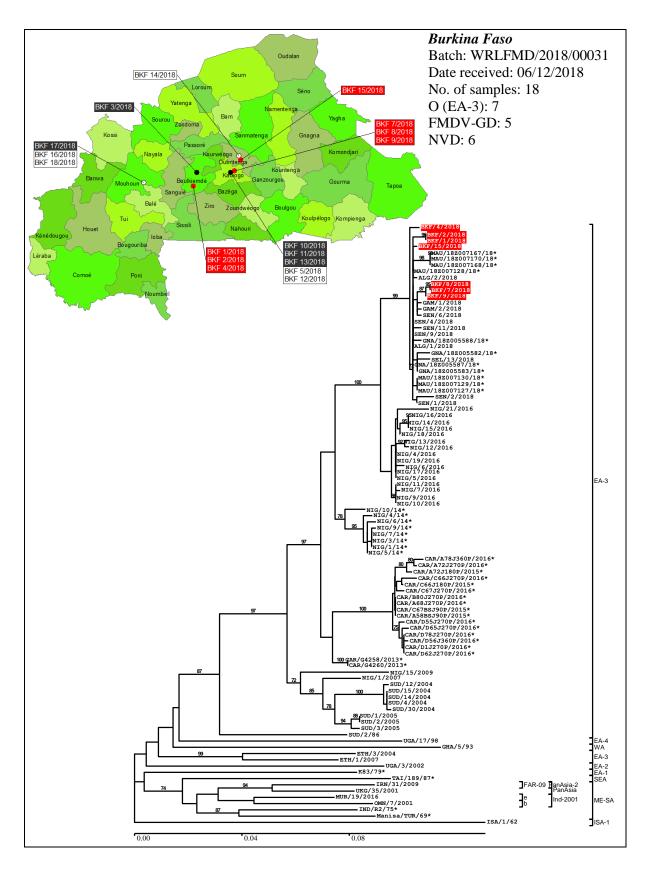




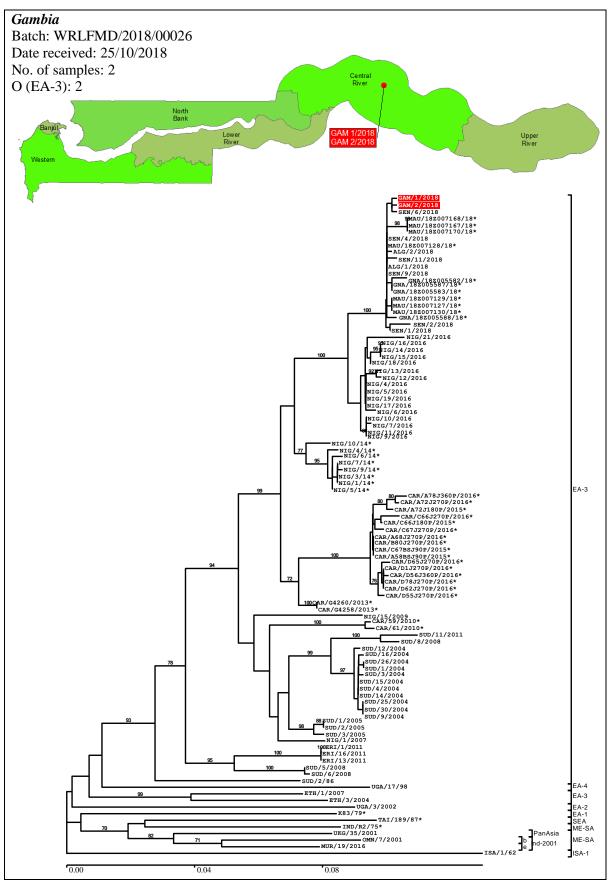




2.2. Africa

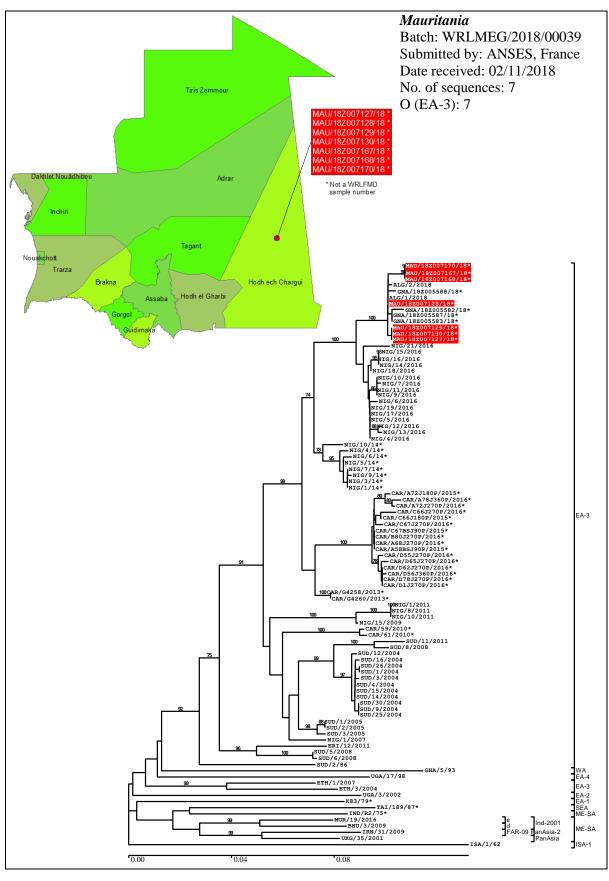




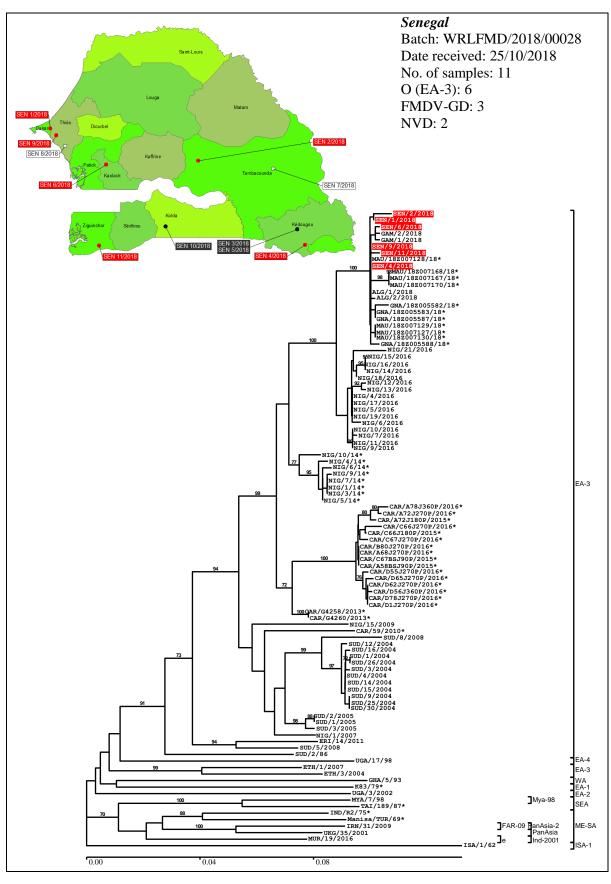


Copies of all the individual reports cited herein can be obtained from The Pirbright Institute and prior to presentation, publication or any other public use of these data, please contact Dr Donald King, The Pirbright Institute, donald.king@pirbright.ac.uk

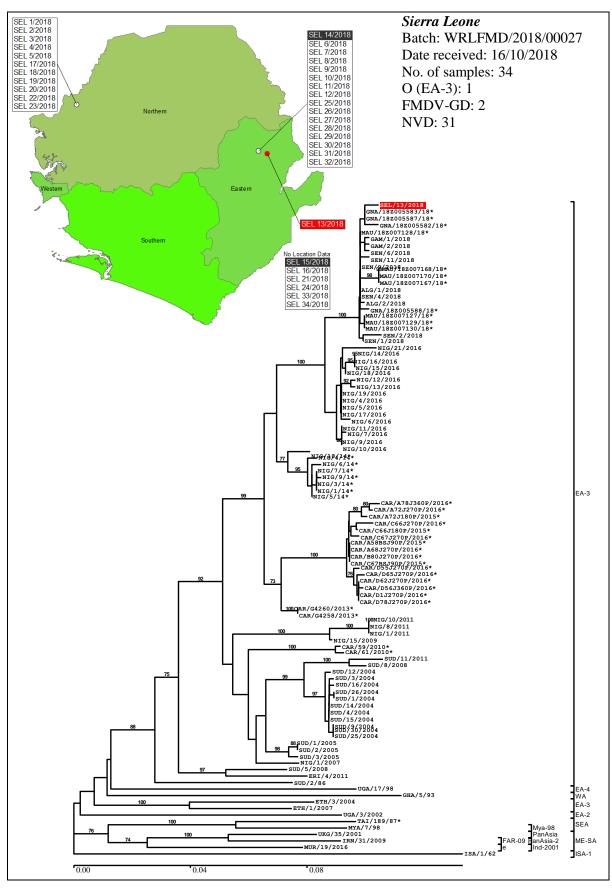




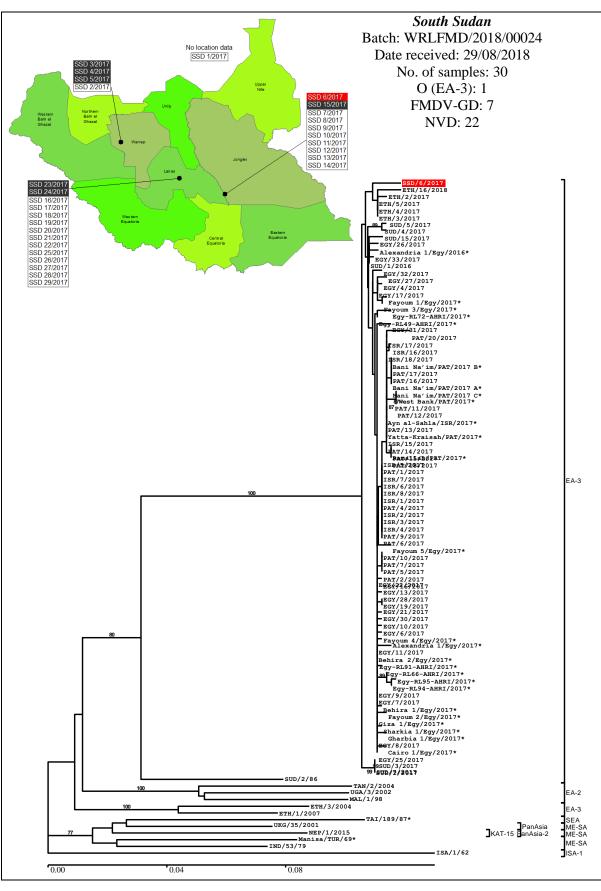














2.3. Vaccine matching

During this reporting period vaccine matching has been undertaken for 28 FMD virus field strains by the WRLFMD® October to December 2018:

Table 3: Summary of samples tested by vaccine matching.

Serotype	0	Α	С	Asia-1	SAT 1	SAT 2	SAT 3
Gambia	1	-	-	-	-	-	-
Hong Kong	1	-	-	-	-	-	-
Kenya	2	2	-	-	1	1	-
Mongolia	3	-	-	-	-	-	-
Senegal	2	-	-	-	-	-	-
Σ	9	2	-	-	1	1	-

For individual data see Annex 1, section 2.6 (Antigenic Characterisation).



Annex 1: Sample data

2.4. Summary of Submissions

Table 4: Summary of samples collected and received to WRLFMD (October to December 2018)

	_	Virus isolation in cell culture/ELISA						RT-PCR for FMD			
Country	Nº of samples	FMD virus serotypes						o Virus etected	(or SVD) virus (where appropriate)		
		0	Α	С	SAT 1	SAT 2	SAT 3	ASIA -1	No V	Danisina	-
BURKINA FASO	18	7	-	-	-	-	-	-	11	12	6
GAMBIA	2	2	-	-	-	-	-	-	-	1	1
HONG KONG	8	1	-	-	-	-	-	-	1	3	5
LAOS	1	-	-	-	-	-	-	-	1	1	-
SENEGAL	11	6	-	-	-	-	-	-	5	8	3
SIERRA LEONE	34	-	-	-	-	-	-	-	34	3	31
THAILAND	19	8	8	-	-	-	-	-	3	19	-
TOTAL	93	24	8	-	-	-	-	-	55	47	46

Abbreviations used in table

VI / ELISA	FMD (or SVD) virus serotype identified following virus isolation in cell culture and antigen detection ELISA
FMD	Foot-and-mouth disease
SVD	Swine vesicular disease
NVD	No FMD, SVD or vesicular stomatitis virus detected
NT	Not tested
rRT-PCR	Real-time reverse transcription polymerase chain reaction for FMD (or SVD) viral genome



2.5. Clinical Samples

Table 5: Clinical sample diagnostics made by the WRLFMD® October to December 2018

	Da	ate					Resu	ılts
Country	Received	Reported	WRL for FMD Sample Identification	Animal	Date of Collection	VI/ELISA	RT-PCR	Final report
BURKINA FASO	06-Dec-18	18-Dec-18	BKF 1/2018 BKF 2/2018 BKF 3/2018 BKF 4/2018 BKF 5/2018 BKF 6/2018 BKF 7/2018 BKF 9/2018 BKF 10/2018 BKF 11/2018 BKF 11/2018 BKF 12/2018 BKF 13/2018 BKF 14/2018 BKF 15/2018 BKF 15/2018 BKF 16/2018	CATTLE	02-Jun-18 02-Jun-18 02-Jun-18 02-Jun-18 06-Jul-18 06-Jul-18 06-Jul-18 06-Jul-18 06-Jul-18 06-Jul-18 06-Jul-18 06-Jul-18 18-Jul-18 18-Jul-18 18-Jul-18 07-Aug-18	O O NEG	POS POS POS NEG POS POS POS POS NEG POS NEG POS NEG	O O FMDV GD O NVD NVD O O FMDV GD FMDV GD NVD FMDV GD NVD FMDV GD NVD FMDV GD NVD O NVD FMDV GD
GAMBIA	25- Oct-	14- Nov	GAM 1/2018 GAM 2/2018	CATTLE CATTLE	23-Jul-18 23-Jul-18	0	POS NEG	0
HONG KONG, SAR of PRC	14-Sep-18	01-Oct-18	HKN 10/2018 HKN 11/2018 HKN 12/2018 HKN 13/2018 HKN 14/2018 HKN 15/2018 HKN 16/2018	PIG PIG PIG PIG PIG PIG	07-Jun-18 07-Jun-18 09-Jul-18 16-Jul-18 16-Jul-18 16-Jul-18 17-Jul-18	NEG FMD FMD, O FMD FMD FMD	POS POS NEG POS NEG NEG NEG	FMDV GD FMD FMD FMD, O FMD FMD FMD
LAOS	14- Nov -18	06- Dec -18	HKN 17/2018 LAO 1/2018	PIG CATTLE	17-Jul-18 22-Jan-18	FMD NEG	NEG POS	FMD FMDV GD
SENEGAL	25-Oct-18	14-Nov-18	SEN 1/2018 SEN 2/2018 SEN 3/2018 SEN 4/2018 SEN 5/2018 SEN 6/2018 SEN 7/2018 SEN 8/2018 SEN 9/2018 SEN 10/2018 SEN 11/2018	CATTLE PIG CATTLE	29-May-18 28-Jun-18 11-Jul-18 11-Jul-18 11-Jul-18 12-Jul-18 20-Jul-18 20-Jul-18 24-Jul-18	O O NEG O NEG NEG O NEG O	POS POS POS POS POS NEG NEG POS POS	O O FMDV GD O FMDV GD O NVD NVD O FMDV GD O



	Da	te					Resu	ılts
Country	Received	Reported	WRL for FMD Sample Identification	Animal	Date of Collection	VI/ELISA	RT-PCR	Final report
SIERRA LEONE	16-Oct-18	02-Nov-18	SEL 1/2018 SEL 2/2018 SEL 3/2018 SEL 4/2018 SEL 5/2018 SEL 5/2018 SEL 6/2018 SEL 7/2018 SEL 7/2018 SEL 9/2018 SEL 10/2018 SEL 11/2018 SEL 11/2018 SEL 12/2018 SEL 12/2018 SEL 15/2018 SEL 15/2018 SEL 15/2018 SEL 16/2018 SEL 17/2018 SEL 17/2018 SEL 17/2018 SEL 17/2018 SEL 20/2018 SEL 21/2018 SEL 21/2018 SEL 22/2018 SEL 22/2018 SEL 22/2018 SEL 25/2018 SEL 25/2018 SEL 25/2018 SEL 26/2018 SEL 27/2018 SEL 29/2018 SEL 29/2018 SEL 29/2018 SEL 29/2018 SEL 29/2018 SEL 30/2018 SEL 30/2018 SEL 31/2018 SEL 31/2018 SEL 33/2018 SEL 33/2018 SEL 33/2018 SEL 33/2018 SEL 33/2018	CATTLE	28-Aug-18 28-Aug-18 28-Aug-18 28-Aug-18 28-Aug-18 28-Aug-18 29-Aug-18 28-Aug-18 28-Aug-18 28-Aug-18 29-Aug-18	NEEGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGG	NEGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGG	NVD
THAILAND	14-Nov-18	06-Dec-18	TAI 6/2017 TAI 7/2017 TAI 8/2017 TAI 9/2017 TAI 10/2017 TAI 11/2017 TAI 12/2017 TAI 13/2017 TAI 15/2017 TAI 16/2017 TAI 16/2017 TAI 17/2017 TAI 18/2017 TAI 19/2017 TAI 19/2017 TAI 1/2018 TAI 2/2018	CATTLE	22-Feb-17 01-Mar-17 28-Mar-17 24-Apr-17 26-Apr-17 10-Nov-17 17-Nov-17 20-Nov-17 23-Nov-17 15-Dec-17 18-Dec-17 25-Dec-17 27-Dec-17 09-Mar-18	A A A A A O O NEG O O A O A G NEG	POS POS POS POS POS POS POS POS POS POS	A A A A A O O FMDV GD O A O A FMDV GD FMDV GD FMDV GD



	Da	ite				Results			
Country	WRL for FMD Sample Anir Oldentification			Animal	Date of Collection	VI/ELISA	RT-PCR	Final report	
		-	TAI 3/2018	CATTLE	09-Jul-18	0	POS	0	
			TAI 4/2018	CATTLE	09-Jul-18	0	POS	0	
			TAI 5/2018	CATTLE	08-Aug-18	0	POS	0	
			TOTAL	93	_				

Abbreviations used in table

FMD(V)	Foot-and-mouth disease (virus)
FMDV GD	Genome detected
FMDV NGD	Genome not detected (samples submitted in Trizol, only rRT-PCR carried out)
VI/ELISA	FMDV serotype identified following virus isolation in cell culture and antigen ELISA
rRT-PCR	Real-time reverse transcription polymerase chain reaction on epithelial suspension for FMD (or SVD) viral genome
NVD	No foot-and-mouth disease, swine vesicular disease or vesicular stomatitis virus detected
NT	Not tested



2.6. Antigenic Characterisation

Antigenic characterisation of FMD field isolates by matching with vaccine strains by 2dmVNT from October to December 2018.

Table 6: Vaccine matching studies for O FMDV by VNT

Strain	Serotype	Topotype	Lineage	O 3039	O1 Manisa	O/TUR/5/2009
MOG/02/2018	0	ME-SA	Ind-2001e	0.55	0.37	0.66
MOG/07/2018	0	SEA	Mya-98	0.40	0.19	0.46
MOG/10/2018	0	ME-SA	PanAsia	0.62	0.44	0.51
HKN/13/2018	0	CATHAY	-	0.11	0.09	0.11
KEN/11/2017	0	EA-2	-	1	1	0.81
KEN/15/2017	0	EA-2	-	0.5	0.52	0.47
GAM/01/2018	0	EA-3	-	0.45	0.30	0.52
SEN/02/2018	0	EA-3	-	0.38	0.30	0.49
SEN/11/2018	0	EA-3	-	0.63	0.39	0.54

Table 7: Vaccine matching studies for A FMDV by VNT

Strain	Serotype	Topotype	Lineage	A/IRN/05	A/TUR/20/06	A22 IRAQ	A/ERI/3/98
KEN/14/2017	Α	AFRICA	G-I	0.05	0.00	0.21	0.20
KEN/17/2017	Α	AFRICA	G-I	0.03	0.00	0.30	0.10



Table 8: Vaccine matching studies for SAT 1 FMDV by VNT

Strain	Serotype	Topotype	Lineage	SAT 1/RHO/12/78
KEN/08/2017	SAT 1	I (NWZ)	-	0.25

Table 9: Vaccine matching studies for SAT 2 FMDV by VNT

Strain	Serotype	Topotype	Lineage	SAT 2 ERI	SAT 2 ZIM
KEN/19/2017	SAT 2	IV	-	0.50	0.31



Abbreviations used in tables

М	Vaccine Match $r_1 = \ge 0.3$. Suggests that there is a close relationship between field isolate and vaccine strain. A potent vaccine containing the vaccine strain is likely to confer protection.
N	No Vaccine Match $r_1 = < 0.3$. Suggests that the field isolate is so different from the vaccine strain that the vaccine is unlikely to protect
В	Borderline Any r ₁ values between 0.28 to 0.32
NT	Not tested against this vaccine



Annex 2: FMD publications

Recent FMD Publications (October to December 2018) cited by Web of Science (Pirbright Institute papers and authors are highlighted in **BOLD AND GREY**)

- 1. Abdrakhmanov, S.K., S.B. Tyulegenov, F.I. Korennoy, A.A. Sultanov, Sytnik, II, K.K. Beisembaev, A.A. Bainiyazov, A.E. Munsey, A.M. Perez, and K. VanderWaal (2018). Spatiotemporal analysis of foot-and-mouth disease outbreaks in the Republic of Kazakhstan, 1955-2013. *Transboundary and Emerging Diseases*, **65**(5): 1235-1245.
- 2. Adhikari, G., K.P. Acharya, M. Upadhyay, R. Raut, K. Kaphle, T. Khanal, M.R. Bertram, C. Stenfeldt, and J. Arzt (2018). Outbreak investigations of *Foot-and-Mouth Disease virus* in Nepal between 2010 and 2015 in the context of historical serotype occurrence. *Veterinary Medicine and Science*, **4**(4): 304-314.
- 3. Ali, W., M. Habib, and M. Shah (2018). Novel strategy for the rapid detection and characterization *Foot-and-Mouth Disease virus* (FMDV) serotypes O, A, and Asia-1. *Journal of Animal and Plant Sciences*, **28**(5): 1301-+.
- 4. Armson, B., V. Mioulet, C. Doel, M. Madi, S. Parida, K.A. Lemire, D.J. Holder, A. Das, M.T. McIntosh, and D.P. King (2018). Detection of Foot-and-Mouth Disease Virus in milk samples by real-time reverse transcription polymerase chain reaction: Optimisation and evaluation of a high-throughput screening method with potential for disease surveillance. Veterinary Microbiology, 223: 189-194.
- 5. Arzt, J., G.J. Belsham, L. Lohse, A. Botner, and C. Stenfeldt (2018). Transmission of Foot-and-Mouth Disease from Persistently Infected Carrier Cattle to Naive Cattle via Transfer of Oropharyngeal Fluid. *Msphere*, **3**(5): 12.
- 6. Bachanek-Bankowska, K., A. Di Nardo, J. Wadsworth, V. Mioulet, G. Pezzoni, S. Grazioli, E. Brocchi, S.C. Kafle, R. Hettiarachchi, P.L. Kumarawadu, I.M. Eldaghayes, A.S. Dayhum, D. Meenowa, S. Sghaier, H. Madani, N. Abouchoaib, B.H. Hoang, P.P. Vu, K. Dukpa, R.B. Gurung, S. Tenzin, U. Wernery, A. Panthumart, K.B. Seeyo, W. Linchongsubongkoch, A. Relmy, L. Bakkali-Kassimi, A. Scherbakov, D.P. King, and N.J. Knowles (2018). Reconstructing the evolutionary history of pandemic foot-and-mouth disease viruses: the impact of recombination within the emerging O/ME-SA/Ind-2001 lineage. Scientific Reports, 8: 11.
- 7. Barrera, J., D.A. Brake, C. Schutta, D. Ettyreddy, B.J. Kamicker, M.V. Rasmussen, C.B. de Rueda, M. Zurita, M. Pisano, W. Hurtle, D.E. Brough, B.T. Butman, B.G. Harper, and J.G. Neilan (2018). Versatility of the adenovirus-vectored foot-and-mouth disease vaccine platform across multiple *Foot-and-Mouth Disease Virus* serotypes and topotypes using a vaccine dose representative of the AdtA24 conditionally licensed vaccine. *Vaccine*, **36**(48): 7345-7352.



- 8. Bertram, M.R., C.B. de Rueda, R. Garabed, S.D. Jumbo, M. Moritz, S. Pauszek, S. Abdoulkadiri, L.L. Rodriguez, and J. Arzt (2018). Molecular Epidemiology of *Foot-and-Mouth Disease Virus* in the Context of Transboundary Animal Movement in the Far North Region of Cameroon. *Frontiers in Veterinary Science*, **5**: 14.
- 9. Cabezas, A.H., M.W. Sanderson, M. Jaberi-Douraki, and V.V. Volkova (2018). Clinical and infection dynamics of foot-and-mouth disease in beef feedlot cattle: An expert survey. *Preventive Veterinary Medicine*, **158**: 160-168.
- Cao, Y.M., W. Zhou, X.C. Xing, J. Zhang, Y.F. Fu, K. Li, P. Sun, P.H. Li, X.W. Bai, X.Q. Ma, H.F. Bao, D. Li, Y.L. Chen, Z.J. Lu, and Z.X. Liu (2018). Indirect ELISA using a multi-epitope recombinant protein to detect antibodies against Foot-and-Mouth Disease Virus serotype O in pigs. Journal of Virological Methods, 262: 26-31.
- 11. Chaters, G., J. Rushton, T.D. Dulu, and **N.A. Lyons** (2018). Impact of foot-and-mouth disease on fertility performance in a large dairy herd in Kenya. *Preventive Veterinary Medicine*, **159**: 57-64.
- 12. Chen, J., X.M. Yu, Q.S. Zheng, L.T. Hou, L.P. Du, Y.P. Zhang, X.W. Qiao, J.B. Hou, and K.H. Huang (2018). The immunopotentiator CVC1302 enhances immune efficacy and protective ability of *Foot-and-Mouth Disease Virus* vaccine in pigs. *Vaccine*, **36**(52): 7929-7935.
- 13. Cima, G. (2018). AVMA wants expanded FMD vaccine bank. *Javma-Journal of the American Veterinary Medical Association*, **253**(5): 530-530.
- 14. de la Higuera, I., C. Ferrer-Orta, E. Moreno, A.I. de Avila, M.E. Soria, K. Singh, F. Caridi, F. Sobrino, S.G. Sarafianos, C. Perales, N. Verdaguer, and E. Domingo (2018). Contribution of a Multifunctional Polymerase Region of Foot-and-Mouth Disease Virus to Lethal Mutagenesis. Journal of Virology, 92(20): 17.
- 15. Defaus, S., M. Forner, L. Hoh, F. Sobrino, E. Blanco, and D. Andreu (2018). Expanding the potential and multivalency of the B2T synthetic peptide vaccine against *Foot-and-Mouth Disease Virus*. *Journal of Peptide Science*, **24**: S57-S57.
- 16. Diez, M., M. Trotta, V. Alfonso, O. Taboga, and M.G. Lopez (2018). Recombinant occlusion bodies of baculovirus as carriers of a non-structural protein of *Foot-and-Mouth Disease Virus*. *3 Biotech*, **8**(11): 6.
- 17. Farooq, U., Z. Ahmed, K. Naeem, M. Bertram, B. Brito, C. Stenfeldt, S.J. Pauszek, M. LaRocco, L. Rodriguez, and J. Arzt (2018). Characterization of naturally occurring, new and persistent subclinical foot-and-mouth disease virus infection in vaccinated Asian buffalo in Islamabad Capital Territory, Pakistan. *Transboundary and Emerging Diseases*, **65**(6): 1836-1850.
- 18. Feng, H.H., Z.X. Zhu, W.J. Cao, F. Yang, X.L. Zhang, X.L. Du, K.S. Zhang, X.T. Liu, and H.X. Zheng (2018). *Foot-and-Mouth Disease Virus* induces lysosomal degradation of NME1 to impair p53-regulated interferon-inducible antiviral genes expression. *Cell Death & Disease*, **9**: 16.



- 19. Fukai, K., T. Nishi, N. Shimada, K. Morioka, M. Yamada, K. Yoshida, and M. Yamakawa (2018). Comparative evaluation of two ELISA kits for detecting antibodies to a nonstructural protein of *Foot-and-Mouth Disease Virus* using serum samples collected from naturally and experimentally infected cows. *Journal of Veterinary Medical Science*, **80**(10): 1624-1630.
- 20. Ganji, V.K., J.K. Biswal, H. Lalzampuia, S.H. Basagoudanavar, P. Saravanan, R.P.T. Selvan, V. Umapathi, G.R. Reddy, A. Sanyal, and H.J. Dechamma (2018). Mutation in the VP2 gene of P1-2A capsid protein increases the thermostability of virus-like particles of *Foot-and-Mouth Disease Virus* serotype O. *Applied Microbiology and Biotechnology*, **102**(20): 8883-8893.
- 21. Gao, F.S., L. Feng, P. Jiang, Z.B. Li, H. Gao, X.X. Zhai, Z.H. Zhang, and X. Hu (2018). Expression, purification, crystallization and preliminary X-ray diffraction analysis of swine leukocyte antigen 2 complexed with a CTL epitope AS64 derived from Asia1 serotype of *Foot-and-Mouth Disease Virus*. *BMC Veterinary Research*, **14**: 6.
- 22. Gelkop, S., A. Sobarzo, P. Brangel, C. Vincke, E. Romao, S. Fedida-Metula, N. Strom, I. Ataliba, F.N. Mwiine, S. Ochwo, L. Velazquez-Salinas, R.A. McKendry, S. Muyldermans, J.J. Lutwama, E. Rieder, V. Yavelsky, and L. Lobel (2018). The Development and Validation of a Novel Nanobody-Based Competitive ELISA for the Detection of Foot-and-Mouth Disease 3ABC Antibodies in Cattle. Frontiers in Veterinary Science, 5: 13.
- 23. Gillies, M. (2018). Modernizing Canada's foot-and-mouth disease response plan. *Canadian Veterinary Journal*, **59**(8): 899-902.
- 24. Gladue, D.P., E. Largo, I. de la Arada, V.M. Aguilella, A. Alcaraz, J.L.R. Arrondo, L.G. Holinka, E. Brocchi, E. Ramirez-Medina, E.A. Vuono, K.A. Berggren, C. Carrillo, J.L. Nieva, and M.V. Borca (2018). Molecular Characterization of the Viroporin Function of *Foot-and-Mouth Disease Virus* Nonstructural Protein 2B. *Journal of Virology*, **92**(23): 19.
- 25. Hagerman, A.D., D.D. South, T.C. Sondgerath, K.A. Patyk, R.L. Sanson, R.S. Schumacher, A.H. Delgado, and S. Magzamen (2018). Temporal and geographic distribution of weather conditions favorable to Check 1 airborne spread of foot-and-mouth disease in the coterminous United States. *Preventive Veterinary Medicine*, **161**: 41-49.
- 26. Horsington, J., C. Nfon, J.L. Gonzales, N. Singanallur, H. Bittner, and W. Vosloo (2018). Protection in sheep against heterologous challenge with serotype Asia-1 *Foot-and-Mouth Disease Virus* using high potency vaccine. *Vaccine*, **36**(41): 6095-6102.
- 27. Jaworski, J.P., J.M. Sala, and A. Capozzo (2018). *Bovine leukemia virus* infection in adult cows does not interfere with foot-and-mouth disease vaccination. *Journal of Dairy Science*, **101**(12): 11247-11250.
- 28. Kang, Y.L., J.Y. Jeong, H.Y. Choi, Y. Zhang, Y. Liu, H.J. Lee, J.C. Choi, S.H. Lee, B.J. Lee, S.W. Lee, J.B. Lee, K.H. Cho, and S.Y. Park (2018). Evaluation and optimization of a conventional SPCE for FMD post-vaccination monitoring. *BMC Veterinary Research*, **14**: 8.



- 29. Kerfua, S.D., G. Shirima, L. Kusiluka, C. Ayebazibwe, R. Mwebe, S. Cleaveland, and D. Haydon (2018). Spatial and temporal distribution of footand-mouth disease in four districts situated along the Uganda-Tanzania border: Implications for cross-border efforts in disease control (vol 85, a1528, 2018). Onderstepoort Journal of Veterinary Research, **85**(1): 1.
- 30. Kinsley, A.C., K. VanderWaal, M.E. Craft, R.B. Morrison, and A.M. Perez (2018). Managing complexity: Simplifying assumptions of foot-and-mouth disease models for swine. *Transboundary and Emerging Diseases*, **65**(5): 1307-1317.
- 31. Kouato, B.S., K. De Clercq, E. Abatih, F. Dal Pozzo, **D.P. King**, E. Thys, H. Marichatou, and C. Saegerman (2018). Review of epidemiological risk models for foot-and-mouth disease: Implications for prevention strategies with a focus on Africa. *Plos One*, **13**(12): 22.
- 32. Kristensen, T., **J. Newman**, S.H. Guan, **T.J. Tuthill**, and G.J. Belsham (2018). Cleavages at the three junctions within the *Foot-and-Mouth Disease Virus* capsid precursor (P1-2A) by the 3C protease are mutually independent. *Virology*, **522**: 260-270.
- 33. Lazarus, D.D., O.L. van Schalkwyk, R.E.J. Burroughs, A. Mpehle, B. Reininghaus, O. Rikhotso, L. Heath, F.F. Maree, B. Blignaut, and G.T. Fosgate (2018). Serological responses of cattle inoculated with inactivated trivalent foot-and-mouth disease vaccine at the wildlife-livestock interface of the Kruger National Park, South Africa. *Preventive Veterinary Medicine*, **158**: 89-96.
- 34. Li, M., T. Xin, X.T. Gao, J. Wu, X.X. Wang, L.C. Fang, X.K. Sui, H.F. Zhu, S.J. Cui, and X.Y. Guo (2018). *Foot-and-Mouth Disease Virus* non-structural protein 2B negatively regulates the RLR-mediated IFN-β induction. *Biochemical and Biophysical Research Communications*, **504**(1): 238-244.
- 35. Li, X., X.P. Meng, S.N. Wang, Z.Q. Li, L. Yang, L.Q. Tu, W.Z. Diao, C. Yu, Y.L. Yu, C.Y. Yan, and L.Y. Wang (2018). Virus-like particles of recombinant PCV2b carrying FMDV-VP1 epitopes induce both anti-PCV and anti-FMDV antibody responses. *Applied Microbiology and Biotechnology*, **102**(24): 10541-10550.
- 36. Lim, D.R., H.R. Kim, M.J. Park, H.G. Chae, B.K. Ku, J.J. Nah, S. Ryoo, S.H. Wee, and C.K. Park (2018). A tailored reverse transcription loop-mediated isothermal amplification for sensitive and specific detection of serotype A *Foot-and-Mouth Disease Virus* circulating in pool 1 region countries. *Transboundary and Emerging Diseases*, **65**(6): 1898-1908.
- 37. Lim, D.R., H.R. Kim, M.J. Park, H.G. Chae, B.K. Ku, J.J. Nah, S.Y. Ryoo, S.H. Wee, Y.R. Park, H.S. Jeon, J.J. Kim, B.Y. Jeon, H.W. Lee, S.G. Yeo, and C.K. Park (2018). An improved reverse transcription loop-mediated isothermal amplification assay for sensitive and specific detection of serotype O *Foot-and-Mouth Disease Virus*. *Journal of Virological Methods*, **260**: 6-13.



- 38. Mansour, K.A., H.H. Naser, and M.H. Hussain (2018). Clinical, molecular detection and phylogenetic analysis study of local *Foot-and-Mouth Disease Virus* in Al-Qadisiyah province of Iraq. *Veterinary World*, **11**(9): 1210-1213.
- 39. Medina, G.N., F. Diaz-San Segundo, C. Stenfeldt, J. Arzt, and T. de los Santos (2018). The Different Tactics of *Foot-and-Mouth Disease Virus* to Evade Innate Immunity. *Frontiers in Microbiology*, **9**: 22.
- 40. Miller, C.A.J., J.R. Young, S. Nampanya, S. Khounsy, N.B. Singanallur, W. Vosloo, R. Abila, S.A. Hamilton, R.D. Bush, and P.A. Windsor (2018). Risk factors for emergence of exotic foot-and-mouth disease O/ME-SA/Ind-2001d on smallholder farms in the Greater Mekong Subregion. *Preventive Veterinary Medicine*, **159**: 115-122.
- 41. Nampanya, S., S. Khounsy, R. Abila, and P.A. Windsor (2018). Implementing large Foot and Mouth Disease vaccination programmes for smallholder farmers: lessons from Lao PDR. *Epidemiology and Infection*, **146**(16): 2086-2095.
- 42. Naveed, A., S.U. Rahman, M.I. Arshad, and B. Aslam (2018). Immune Modulatory Potential of Anti-idiotype Antibodies as a Surrogate of *Foot-and-Mouth Disease Virus* Antigen. *Msphere*, **3**(5): 10.
- 43. Railey, A.F., T. Lembo, G.H. Palmer, G.M. Shirima, and T.L. Marsh (2018). Spatial and temporal risk as drivers for adoption of foot-and-mouth disease vaccination. *Vaccine*, **36**(33): 5077-5083.
- 44. Raut, N., J. Riviere, S. Hosteing, E. Collin, S. Philizot, O. Debaere, and G. Zanella (2018). Overview of foot-and-mouth disease awareness among farmers and veterinarians in France. *Veterinary Record*, **183**(5): 161-+.
- 45. Sharma, A.K., M. Bhatt, M. Sankar, J.K. Mohapatra, B.B. Dash, G.R. Gowane, S. Subramaniam, R. Ranjan, and B. Pattnaik (2018). Kinetics of Interferon gamma and Interleukin-21 response following *Foot-and-mouth disease virus* infection. *Microbial Pathogenesis*, **125**: 20-25.
- 46. Teng, Z.D., S.Q. Sun, H. Chen, J. Huang, P. Du, H. Dong, X.Y. Xu, S.Y. Mu, Z.J. Zhang, and H.C. Guo (2018). Golden-star nanoparticles as adjuvant effectively promotes immune response to *Foot-and-mouth disease virus*-like particles vaccine. *Vaccine*, **36**(45): 6752-6760.
- 47. Veerapen, V.P., A.R. van Zyl, E.P. Rybicki, and A.E. Meyers (2018). Transient expression of heat- and acid-resistant *Foot-and-mouth disease virus* P1-2A mutants in *Nicotiana benthamiana*. *Virus Research*, **256**: 45-49.
- 48. Wang, H.M., P.L. Hou, G.M. Zhao, L. Yu, Y.W. Gao, and H.B. He (2018). Development and evaluation of serotype-specific recombinase polymerase amplification combined with lateral flow dipstick assays for the diagnosis of *Foot-and-mouth disease virus* serotype A, O and Asia1. *BMC Veterinary Research*, **14**: 11.
- 49. Wang, H.M., G.M. Zhao, P.L. Hou, L. Yu, C.Q. He, and H.B. He (2018). Rapid detection of *Foot-and-mouth disease virus* using reverse transcription



- recombinase polymerase amplification combined with a lateral flow dipstick. *Journal of Virological Methods*, **261**: 46-50.
- 50. Wiratsudakul, A. and S. Sekiguchi (2018). The implementation of cattle market closure strategies to mitigate the foot-and-mouth disease epidemics: A contact modeling approach. *Research in Veterinary Science*, **121**: 76-84.
- 51. Wu, B., H. Zhang, K. Li, K. Mehmood, Y. Zhao, B. Jiang, C.J. Xue, M.T. Javed, F. Nabi, Z.Q. Han, and H.Q. Luo (2018). Seroprevalence and Immunization Program of Serotype O of *Foot-and-Mouth Disease Virus* in Pigs in Zhejiang Province, China. *Pakistan Journal of Zoology*, **50**(5): 1945-1949.
- 52. Zhu, Z., F. Yang, J. He, J. Li, W. Cao, J. Li, Y. Xia, J. Guo, Y. Jin, K. Zhang, H. Zheng, and X. Liu (2018). First detection of *Foot-and-mouth disease virus* O/ME-SA/Ind2001 in China. *Transboundary and Emerging Diseases*, **65**(6): 2027-2031.
- 53. Zhu, Z.X., X.L. Du, P.F. Li, X.L. Zhang, F. Yang, W.J. Cao, H. Tian, K.S. Zhang, X.T. Liu, and H.X. Zheng (2018). Early Growth Response Gene-1 Suppresses *Foot-and-Mouth Disease Virus* Replication by Enhancing Type I Interferon Pathway Signal Transduction. *Frontiers in Microbiology*, **9**: 12.

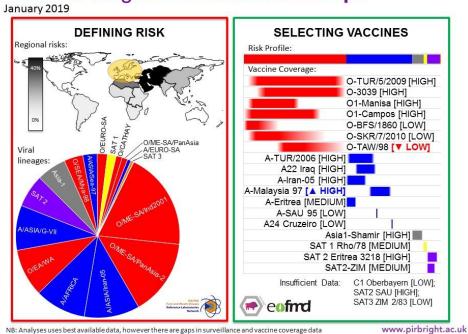


Annex 3: Vaccine Recommendations

This report provides recommendations of FMDV vaccines to be included in antigen banks. These outputs are generated with a new tool (called PRAGMATIST) that has been developed in partnership between WRLFMD® and EuFMD. These analyses accommodate the latest epidemiological data collected by the OIE FAO FMD Laboratory Network regarding FMDV lineages that are present in different *source regions* (see Table below), as well as available *in vitro*, *in vivo* and field data to score the ability of vaccines to protect against these FMDV lineages.

Lineage	West Eurasia	East Asia	North Africa	India and Southern Asia	East Africa	West and Central Africa	Southern Africa	South America
O ME-SA PanAsia-2	35	-	-	-	-	-	-	-
O ME-SA PanAsia	-	10	-	-	-	-	-	-
O SEA Mya-98	-	33	-	-	-	-	-	-
O ME-SA Ind2001	6	20	35	80	-	-	-	-
O EA or O WA	3	-	20	-	45	37	-	-
O EURO-SA	-	-	-	-	-	-	-	74
O CATHAY	-	10.5	-	-	-	-	-	-
A ASIA Sea-97	-	25	-	-	-	-	-	-
A ASIA Iran-05	25.5	-	-	-	-	-	-	-
A ASIA G-VII	17.5	-	-	16	-	-	-	-
A AFRICA	-	-	35	-	24	25	-	-
A EURO-SA	-	-	-	-	-	-	-	26
Asia-1	12.5	1.5	-	4	-	-	-	-
SAT 1	-	-	-	-	10	10	27	-
SAT 2	0.5	-	10	-	20	28	57	-
SAT 3	-	-	-	-	1		16	-
С	-	-	-	-	-	-	-	-

Vaccine Antigen Prioritisation: Europe



The table defines the relative distribution of FMDV lineages in each of the eight *source regions*, while the figure highlights the importance of these *source regions* for **Europe** (using data collected at the EU-RL Workshop); please contact WRLFMD EuFMD for assistance to tailor these outputs to other geographical regions. NB: Vaccine-coverage data presented is based on available data and may under-represent the true performance of individual vaccines.



Annex 4: Other WRLFMD Activities

Proficiency test scheme organised by WRLFMD:

Phase XXX: sample panels are being dispatched to participating laboratories (including "live" and inactivated samples for virology assays, and validated sera for FMDV-specific antibody tests). Please contact WRLFMD if you would like more information about this exercise.

Summary of participating laboratories (status as of January 2019):

	Phase XXX			
Total invited laboratories	102			
Participants from European Union	26			
(funded by EURL for FMD)	(EU member states)			
Participants from Global Network	Argentina, Brazil, Canada, Russia, Senegal, Thailand Pending: Botswana, China, Ethiopia, India, Kenya, Nepal, Nigeria, Republic of Korea, South Africa, USA			
Participants from EuFMD Member states (non-EU)	Bosnia & Herzegovina, Georgia, Kosovo, FYRO Macedonia, Norway, Serbia, Switzerland, Turkey Pending: Albania,			
Participants from neighbourhood countries	Algeria, Armenia, Montenegro, Morocco Pending: Belarus, Iran, Iraq, Jordan, Lebanon, Moldova, Tunisia, Ukraine			
Other participating countries	Australia, Namibia, New Zealand, Singapore, Chinese Taipei,			



Training courses:

20th **February 2019:** E-learning training course covering FMD Diagnostic methods:

FMD Laboratory Investigation Training Course

This course is aimed at those working in national or regional foot-and-mouth disease laboratories and involved in carrying out or managing laboratory testing activities.

Online training

The course covers the full range of activities carried out by FMD laboratories from supervising collection of diagnostic samples through to advanced laboratory testing procedures, biosafety and quality assurance.

Participants will learn to

- Interact with field staff and guide them in collection and submission of appropriate, quality diagnostic samples
 Select appropriate diagnostic tests to detect FMD virus and FMD
- Select appropriate diagnostic tests to detect FMD virus and FMD virus-specific antibodies, and interpret the results of these tests
- Describe the principles of accurate virus detection test methods and assays used for serology
- Outline techniques for further characterization of FMD virus including genomic sequencing and vaccine matching tests
 Explain the importance and basic principles of laboratory Quality
- Assurance
- Explain the key principles of biosecurity and biosafety measures to be carried out in an FMD laboratory

The course involves 14 hours of interactive e-learning content over a four week period

The course provides a unique opportunity to interact with your colleagues in FMD laboratories around the world

A limited number of places are available on this course To apply, send an email to: eufmd-training@fao.org

Dates and more information on the e-learning courses are available at https://eufmdlearning.works





The EuFMD and the World Reference Laboratory for FMD, based at the Pirbright Institute, have partnered to produce the online FMD Laboratory Investigation