



# WRLFMD Quarterly Report October to December 2016

**Reference Laboratory Contract Report** 

Foot-and-Mouth Disease





Department for Environment Food & Rural Affairs





# CONTENTS

1. S	ummary of samples tested and reported FMD outbreaks	3
1.1.	Asia	3
1.2.	Africa	4
1.3.	South America	5
1.4.	Uncharacterised FMD viruses	5
2. D	etailed Analysis	7
2.1.	ASIA	8
2.2.	Africa	19
3. Va	accine matching	21
	nnex 1	
4.1.	Summary of Submissions	22
4.2.	Clinical Samples	22
4.3.	Antigenic Characterisation	28
5. A	nnex 2	30
6. A	nnex 3	36



# 1. Summary of samples tested and reported FMD outbreaks

# 1.1. Asia

# Afghanistan

A batch of 27 samples were received on 13/09/2016. Fourteen were typed as **O** (n=10), **A** (n=1) or **Asia 1** (n=3). FMDV genome was detected in a further 10 samples and 3 were NVD. The type O viruses all belonged to **ME-SA/PanAsia-2<sup>ANT-10</sup>** lineage; the type A to **ASIA/Iran-05<sup>FAR-11</sup>**; and Asia 1 to **ASIA/Sindh-08** (see below).

# Bhutan

A batch of 14 samples was received on 13<sup>th</sup> December. The typing and sequencing results are pending.

## Mongolia

An outbreak of **FMD type A** occurred on 17/07/2016 in cattle at Muu Khoot, 6 Bag, Sumber soum, Govi-Sumber. VP1 sequencing was performed at the FGI-ARRIAH and comparisons at the WRLFMD showed the outbreaks to be caused by viruses belonging to the **A/ASIA/Sea-97** lineage (see below).

## Myanmar

A batch of four samples was received on 14<sup>th</sup> October. No FMD virus could be isolated, but FMDV genome was detected by rRT-PCR in two of the samples. The VP1 coding region was successfully amplified for one sample (from Rakhine state) and the genotype confirmed as **O/ME-SA/Ind-2001d** as had previously been determined in the Regional Reference Laboratory in Pakchong, Thailand (see below).

## People's Republic of China

In November, outbreaks of FMD type O were reported in two different provinces Jiangxi (in pigs) and Xinjiang (in cattle; near to the border with Kyrgyzstan). No genotyping has been reported.



## Russia

An outbreak of **FMD type Asia 1** was reported to have occurred on 16/10/2016 in cattle at Vyshmanovo, Sobinsky, Vladimirskaya Oblast (western Russia). Genotyping data was presented at the Annual Meeting of OIE/FAO Reference Laboratory Network in November 2016.

In November/December three outbreaks of **FMD type O** occurred in cattle and pigs in Zabaykal'skiy kray in the eastern part of the country close to Mongolia and China. VP1 sequencing was performed at the FGI-ARRIAH and comparisons undertaken by WRLFMD showed the outbreaks to be caused by viruses belonging to the **O/ME-SA/Ind-2001d** lineage (see below).

## Saudi Arabia

An outbreak of **FMD type A** was reported to have occurred on 14/10/2016 in cattle and sheep at Alkharj, Alkharj, Ar Riyad. Three samples were submitted to the WRLFMD and **FMDV types A and O** were isolated. The two type A viruses belonged to the ASIA/G-VII lineage while the type O virus belonged to PanAsia-2<sup>ANT-10</sup> (see below).

## Thailand

A batch to 20 samples was received on 14<sup>th</sup> October. Nine were typed as **FMDV O**, five as **FMDV A** and in the remaining six FMDV genome was detected by rRT-PCR. All the **type O** viruses belonged to the **SEA/Mya-98** lineage and the **type A** viruses to the **ASIA/Sea-97** lineage (see below).

On the 10<sup>th</sup> November, the OIE Regional Reference Laboratory submitted two **FMDV O** VP1 sequences for genotyping. Analyses showed that they belonged to the **ME-SA/Ind-2001d** lineage. This is the fourth Southeast Asian country (the others being Myanmar, Laos and Vietnam) to have reported this lineage in the past year.

## Vietnam

A batch of 35 samples was received on 20<sup>th</sup> December. The typing and sequencing results are pending.

# 1.2. Africa

#### Guinea-Bissau

On 5<sup>th</sup> October, FMD was reported in cattle in the Bafatá Region. Laboratory investigations at the Regional Reference Laboratory in Senegal suggested the



involvement of multiple serotypes. **FMD SAT 1** and **SAT 2** viruses were detected in vesicular fluids while antibody assays suggested **serotypes 0** and **A** may be involved. Confirmation of these findings is needed and genotyping of any virus isolates would be useful.

## Mozambique

Two outbreaks of **FMD type SAT 2** were reported in cattle at the end of September and beginning of October in Gaza and Maputo, respectively. No genotyping has been reported.

#### Nigeria

A batch of 25 samples were received on the 19<sup>th</sup> October. Seventeen samples were typed as **FMDV O** and three as **FMDV SAT 1**. FMDV genome was detected in one sample and four were NVD. This is the first time SAT 1 has been confirmed in West Africa since 1981 when it was identified in Nigeria. One **FMD type O** virus was genotyped as WA and the other 16 as EA-3; the three **SAT 1** viruses were genetically distinct from all other SAT 1 sequences in the WRLFMD database and were assigned a new topotype number, X (see below).

#### Zambia

On 08/10/2016 an outbreak due to **FMD (untyped)** was reported at Kaka Veterinary Camp, Mbala, Northern Province.

#### Zimbabwe

Between 8<sup>th</sup> September and 25<sup>th</sup> October 2016, a further 13 outbreaks of **FMD type SAT 2** were reported in cattle in Matabeleland North (n=11) and Midlands (n=2) provinces. No genotyping has been reported.

# **1.3. South America**

No new outbreaks of FMD were reported in the region.

# 1.4. Uncharacterised FMD viruses

A number of outbreaks have occurred where samples have not been sent to the WRLFMD. It is probable that the countries involved have performed their own genetic characterisation; however, through the OIE/FAO laboratory network we would also like to encourage the submission of samples (or complete VP1 sequences) to the WRLFMD.



An up-to-date list and reports of FMD viruses characterised by sequencing can be found at the following website: <u>http://www.wrlfmd.org/fmd\_genotyping/2016.htm</u>.

Results from samples or sequences received at WRLFMD (status of samples being tested) are shown in Table 1 and a complete list of clinical sample diagnostics made by the WRLFMD between October to December 2016 is shown in Annex 1 (Summary of Submissions). A record of all samples received by WRLFMD (October to December 2016) is shown in Annex 1 (Clinical Samples).

<b>Table 1:</b> Status of sequencing of samples or sequences received by the WRLFMD
from October to December 2016 (* indicates samples carried over from the
last quarter)

WRLFMD Batch No.	Date received	Country	Serotype	No. of samples	No. of sequences	Sequencing status
WRLFMD/2016/00029	13/09/2016	Afghanistan	0	10	10	Completed*
			А	1	1	Completed*
			Asia 1	3	3	Completed*
WRLFMD/2016/00031	14/10/2016	Thailand	0	9	9	Completed
			А	5	5	Completed
WRLFMD/2016/00033	14/10/2016	Myanmar	O*	1	1	Completed
WRLFMD/2016/00034	19/10/2016	Nigeria	0	17	17	Completed
			SAT 1	3	3	Completed
WRLFMD/2016/00035	24/10/2016	Saudi Arabia	0	1	1	Completed
			А	2	2	Completed
WRLFMD/2016/00036	13/12/2016	Bhutan	Pending	14	-	Pending
WRLFMD/2016/00037	20/12/2016	Vietnam	Pending	35	-	Pending
WRLMEG/2016/00022	17/07/2016	Mongolia	А	-	3	Completed
WRLMEG/2016/00025	23/112016	Russia	0	-	2	Completed
			Total	101	57	

\*, received during the 3<sup>nd</sup> quarter of 2016



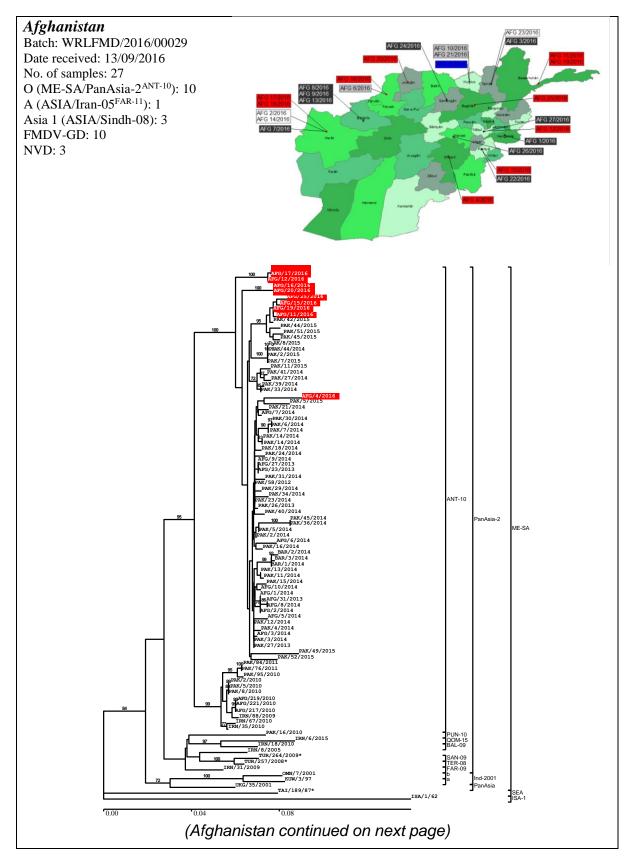
# 2. Detailed Analysis

Key for maps and trees:

Serotype O
Serotype A
Serotype C
Serotype Asia-1
Serotype SAT 1
Serotype SAT 2
Serotype SAT 3
FMDV Genome Detected
No Virus Detected



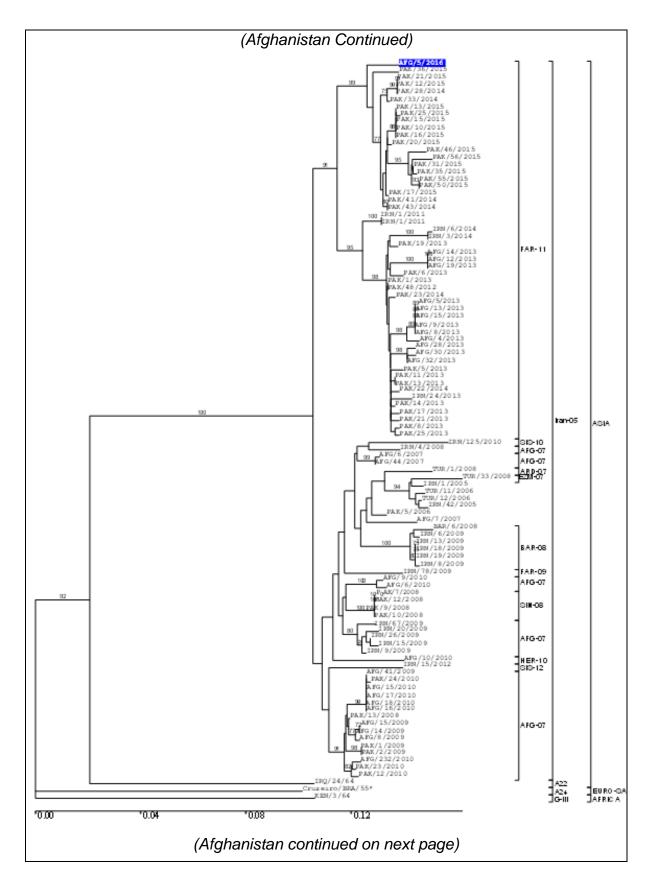
# 2.1. ASIA



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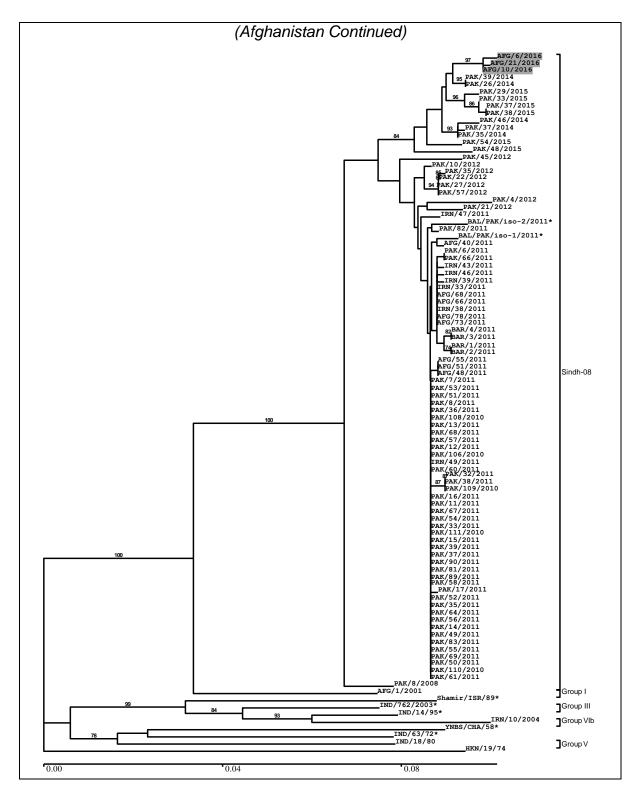
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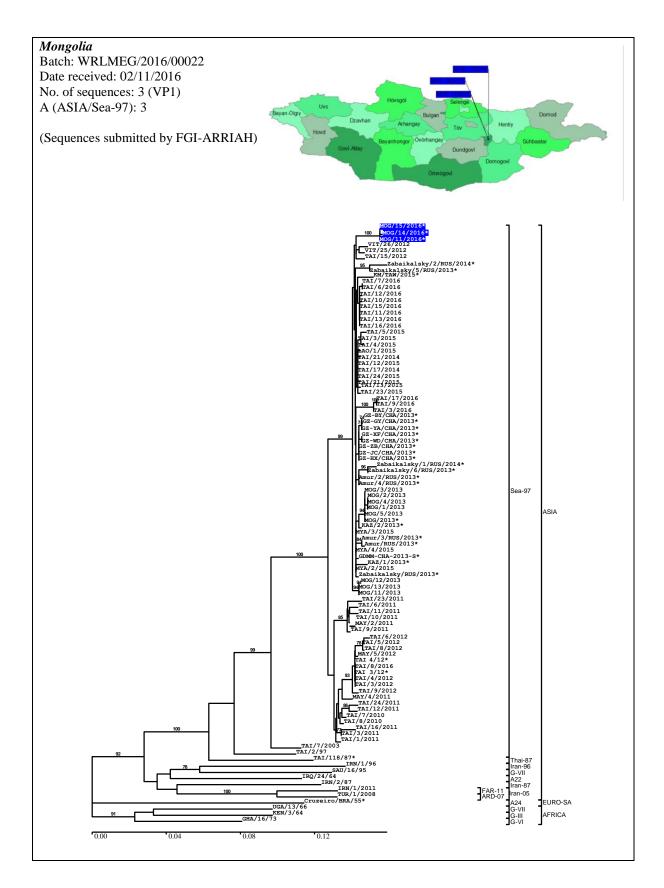


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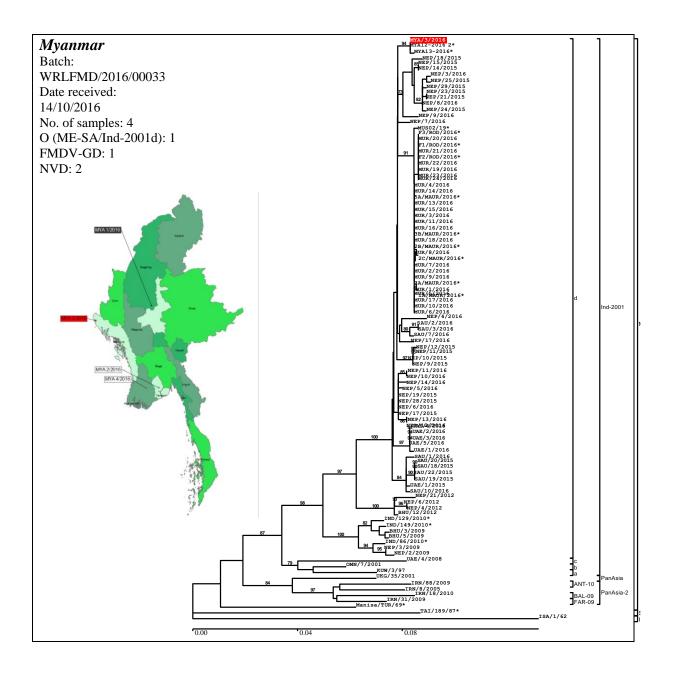




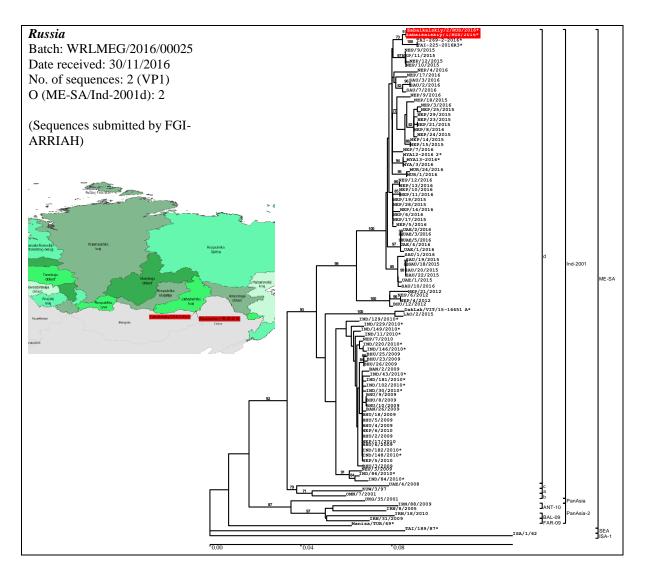


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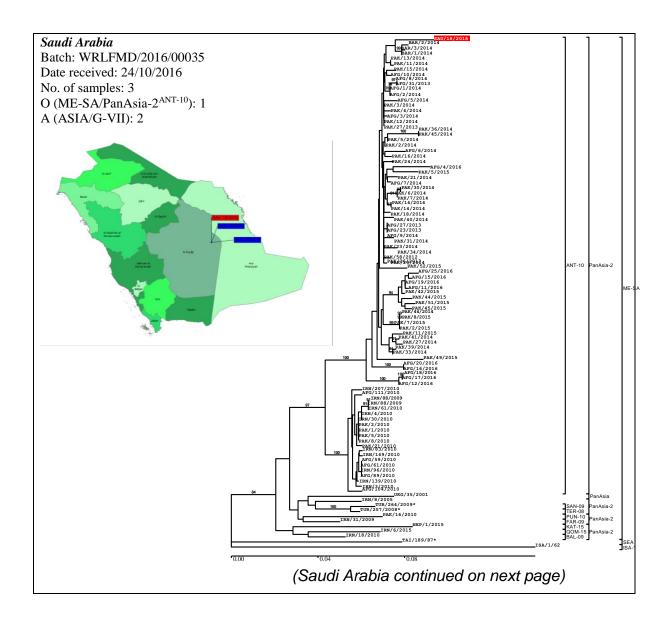








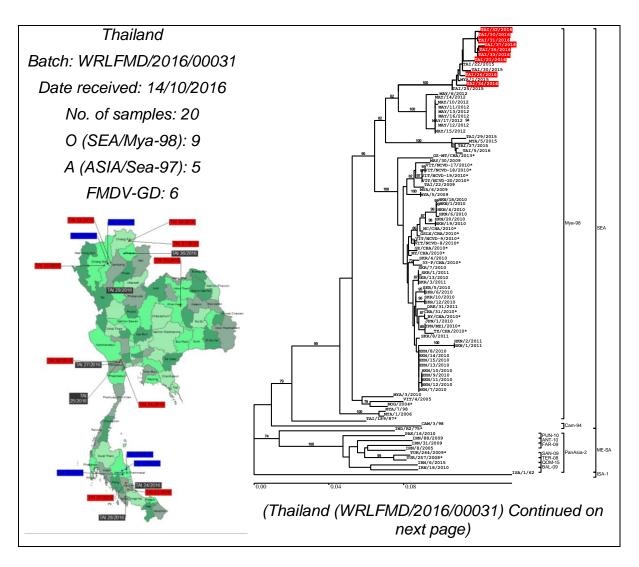




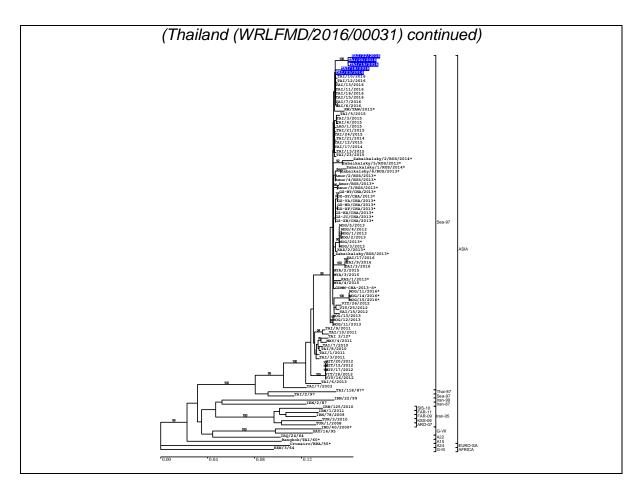




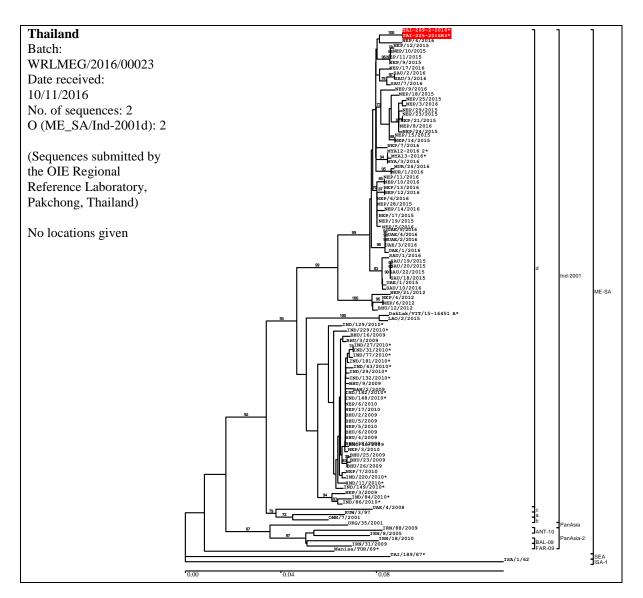






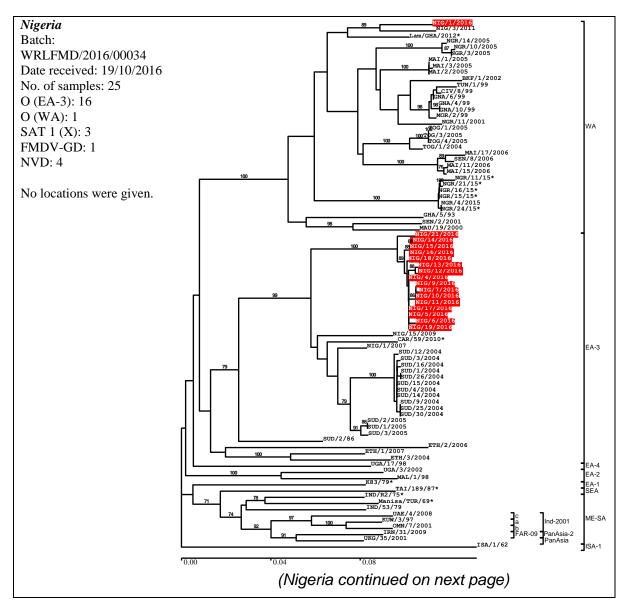




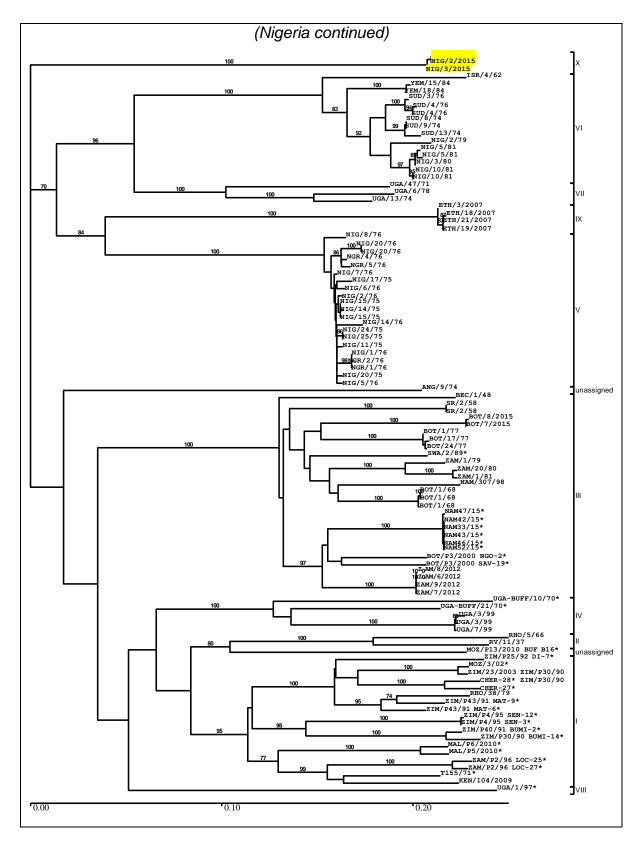




# 2.2. Africa









# 3. Vaccine matching

During this reporting period vaccine matching has been undertaken for 26 FMD virus field strains (serotype O (n=17), serotype A (n=5), serotype Asia 1 (n=2) and serotype SAT 1 (n=2).

For individual data see Annex 1, section 4.3 (Antigenic Characterisation).



# 4. Annex 1

# 4.1. Summary of Submissions

Table 2: Summary of samples collected and received to WRLFMD (October to December 2016)

		Virus isolation in cell culture/ELISA					RT-PCR for FMD (or SVD)				
Country	Nº of samples	FMD virus serotypes					No Virus Detected	virus	(where opriate)		
		0	Α	с	SAT 1	SAT 2	SAT 3	ASIA-1	No	Positive	Negative
BHUTAN	14	-	-	-	-	-	-	-	-	-	-
LAOS	6	-	-	-	-	-	-	-	-	-	-
MYANMAR	4	-	-	-	-	-	-	-	4	2	2
NIGERIA	25	17	-	-	3	-	-	-	5	19	6
SAUDI ARABIA	3	1	2	-	-	-	-	-	-	3	-
THAILAND	20	9	5	-	-	-	-	-	6	20	-
VIETNAM	35	-	-	-	-	-	-	-	-	-	-
TOTAL	107	27	7	-	3	-	-	-	15	44	8

## Abbreviations used in table

VI / ELISA	FMD (or SVD) virus serotype identified following virus isolation in cell culture and antigen detection ELISA
FMD	Foot-and-mouth disease
SVD	Swine vesicular disease
NVD	No FMD, SVD or vesicular stomatitis virus detected
NT	Not tested
rRT-PCR	Real-time reverse transcription polymerase chain reaction for FMD (or SVD) viral genome

# 4.2. Clinical Samples

Table 3: Clinical sample diagnostics made by the WRLFMD® October to December 2016

	WRL for FMD		Date of	Results			
Country	Sample Identification	Animal	Collection	VI/ELISA	RT-PCR	Final report	
BHUTAN	BHU 1/2015	CATTLE	14-Aug-15	Pending	Pending	Pending	
	BHU 2/2015	CATTLE	15-Aug-15	Pending	Pending	Pending	

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0	WRL for FMD	A •	Date of		Results	
Country	Sample Identification	Animal	Collection	VI/ELISA	RT-PCR	Final report
	BHU 3/2015	CATTLE	22-Nov-15	Pending	Pending	Pending
	BHU 4/2015	CATTLE	26-Dec-15	Pending	Pending	Pending
	BHU 1/2016	CATTLE	15-Feb-16	Pending	Pending	Pending
	BHU 2/2016	CATTLE	15-Feb-16	Pending	Pending	Pending
	BHU 3/2016	CATTLE	15-Feb-16	Pending	Pending	Pending
	BHU 4/2016	CATTLE	17-Feb-16	Pending	Pending	Pending
	BHU 5/2016	CATTLE	08-Mar-16	Pending	Pending	Pending
	BHU 6/2016	CATTLE	24-Mar-16	Pending	Pending	Pending
	BHU 7/2016	CATTLE	04-Jun-16	Pending	Pending	Pending
	BHU 8/2016	CATTLE	04-Jun-16	Pending	Pending	Pending
	BUH 9/2016	CATTLE	04-Jun-16	Pending	Pending	Pending
	BHU 10/2016	CATTLE	04-Jun-16	Pending	Pending	Pending
MYANMAR	MYA 1/2016	CATTLE	29-Jun-16	NEG	POS	FMDV GD
	MYA 2/2016	CATTLE	29-Jun-16	NEG	NEG	NVD
	MYA 3/2016	CATTLE	29-Jun-16	NEG	POS	FMDV GD
	MYA 4/2016	CATTLE	29-Jun-16	NEG	NEG	NVD
NIGERIA	NIG 1/2015	BOVINE	02-Dec-15	SAT 1	POS	SAT 1
	NIG 2/2015	BOVINE	02-Dec-15	SAT 1	POS	SAT 1
	NIG 3/2015	BOVINE	02-Dec-15	SAT 1	POS	SAT 1
	NIG 1/2016	BOVINE	27-Jun-16	0	POS	0
	NIG 2/2016	BOVINE	09-Aug-16	NEG	NEG	NVD
	NIG 3/2016	BOVINE	09-Aug-16	NEG	NEG	NVD
	NIG 4/2016	BOVINE	19-Aug-16	0	NEG	0



0	WRL for FMD	A ra :	Date of		Results	
Country	Sample Identification	Animal	Collection	VI/ELISA	RT-PCR	Final report
	NIG 5/2016	BOVINE	19-Aug-16	0	POS	0
	NIG 6/2016	BOVINE	19-Aug-16	0	POS	0
	NIG 7/2016	OVINE	03-Sep-16	0	POS	0
	NIG 8/2016	BOVINE	03-Sep-16	NEG	POS	FMDV GD
	NIG 9/2016	BOVINE	03-Sep-16	0	NEG	0
	NIG 10/2016	BOVINE	03-Sep-16	0	POS	0
	NIG 11/2016	BOVINE	03-Sep-16	0	POS	0
	NIG 12/2016	BOVINE	09-Sep-16	0	POS	0
	NIG 13/2016	BOVINE	09-Sep-16	0	POS	0
	NIG 14/2016	BOVINE	13-Sep-16	0	POS	0
	NIG 15/2016	BOVINE	13-Sep-16	0	POS	0
	NIG 16/2016	BOVINE	13-Sep-16	0	POS	0
	NIG 17/2016	BOVINE	13-Sep-16	0	POS	0
	NIG 18/2016	BOVINE	13-Sep-16	0	POS	0
	NIG 19/2016	BOVINE	13-Sep-16	0	POS	0
	NIG 20/2016	BOVINE	23-Sep-16	NEG	NEG	NVD
	NIG 21/2016	BOVINE	30-Sep-16	0	POS	0
	NIG 22/2016	BOVINE	30-Sep-16	NEG	NEG	NVD
SAUDI ARABIA	SAU 18/2016	CATTLE	14-Oct-16	0	POS	0
	SAU 19/2016	CATTLE	14-Oct-16	А	POS	А
	SAU 20/2016	CATTLE	14-Oct-16	А	POS	А
THAILAND	TAI 18/2016	CATTLE	09-Mar-16	А	POS	А
	TAI 19/2016	CATTLE	18-Mar-16	А	POS	А



0	WRL for FMD	A	Date of		Results	
Country	Sample Identification	Animal	Collection	VI/ELISA	RT-PCR	Final report
	TAI 20/2016	CATTLE	05-Apr-16	А	POS	А
	TAI 21/2016	CATTLE	11-Apr-16	0	POS	0
	TAI 22/2016	CATTLE	18-Apr-16	А	POS	А
	TAI 23/2016	CATTLE	18-Apr-16	А	POS	А
	TAI 24/2016	PIG	17-May-16	NEG	POS	FMDV GD
	TAI 25/2016	PIG	17-May-16	NEG	POS	FMDV GD
	TAI 26/2016	CATTLE	25-May-16	0	POS	0
	TAI 27/2016	CATTLE	08-Jun-16	NEG	POS	FMDV GD
	TAI 28/2016	CATTLE	21-Jul-16	NEG	POS	FMDV GD
	TAI 29/2016	CATTLE	29-Jul-16	NEG	POS	FMDV GD
	TAI 30/2016	CATTLE	29-Jul-16	0	POS	0
	TAI 31/2016	CATTLE	29-Jul-16	0	POS	0
	TAI 32/2016	CATTLE	29-Jul-16	0	POS	0
	TAI 33/2016	PIG	29-Jul-16	0	POS	0
	TAI 34/2016	CATTLE	02-Aug-16	0	POS	0
	TAI 35/2016	CATTLE	02-Aug-16	0	POS	0
	TAI 36/2016	CATTLE	09-Aug-16	NEG	POS	FMDV GD
	TAI 37/2016	CATTLE	11-Aug-16	0	POS	0
VIETNAM	VIT 5/2015	CATTLE	02-Jun-15	Pending	Pending	Pending
	VIT 6/2015	CATTLE	02-Jun-15	Pending	Pending	Pending
	VIT 7/2015	PIG	06-Jun-15	Pending	Pending	Pending
	VIT 8/2015	CATTLE	10-Jul-15	Pending	Pending	Pending
	VIT 9/2015	BUFFALO	14-Sep-15	Pending	Pending	Pending



	WRL for FMD		Date of		Results	
Country	Sample Identification	Animal	Collection	VI/ELISA	RT-PCR	Final report
	VIT 10/2015	CATTLE	14-Sep-15	Pending	Pending	Pending
	VIT 11/2015	PIG	16-Oct-15	Pending	Pending	Pending
	VIT 12/2015	CATTLE	28-Oct-15	Pending	Pending	Pending
	VIT 13/2015	CATTLE	28-Oct-15	Pending	Pending	Pending
	VIT 14/2015	BUFFALO	30-Oct-15	Pending	Pending	Pending
	VIT 15/2015	BUFFALO	10-Nov-15	Pending	Pending	Pending
	VIT 16/2015	CATTLE	24-Nov-15	Pending	Pending	Pending
	VIT 17/2015	PIG	01-Dec-15	Pending	Pending	Pending
	VIT 18/2015	CATTLE	02-Dec-15	Pending	Pending	Pending
	VIT 1/2016	BUFFALO	28-Jan-16	Pending	Pending	Pending
	VIT 2/2016	BUFFALO	16-Feb-16	Pending	Pending	Pending
	VIT 3/2016	CATTLE	29-Feb-16	Pending	Pending	Pending
	VIT 4/2016	PIG	11-Mar-16	Pending	Pending	Pending
	VIT 5/2016	CATTLE	11-Mar-16	Pending	Pending	Pending
	VIT 6/2016	BUFFALO	18-Mar-16	Pending	Pending	Pending
	VIT 7/2016	CATTLE	24-Mar-16	Pending	Pending	Pending
	VIT 8/2016	PIG	28-Mar-16	Pending	Pending	Pending
	VIT 9/2016	CATTLE	06-Apr-16	Pending	Pending	Pending
	VIT 10/2016	CATTLE	12-Apr-16	Pending	Pending	Pending
	VIT 11/2016	PIG	26-Apr-16	Pending	Pending	Pending
	VIT 12/2016	PIG	05-May-16	Pending	Pending	Pending
	VIT 13/2016	CATTLE	15-May-16	Pending	Pending	Pending
	VIT 14/2016	CATTLE	12-Jun-16	Pending	Pending	Pending



	WRL for FMD		Date of	Results			
Country	Sample Identification	Animal	Collection	VI/ELISA	RT-PCR	Final report	
	VIT 15/2016	BUFFALO	03-Aug-16	Pending	Pending	Pending	
	VIT 16/2016	PIG	09-Aug-16	Pending	Pending	Pending	
	VIT 17/2016	CATTLE	25-Aug-16	Pending	Pending	Pending	
	VIT 18/2016	CATTLE	27-Aug-16	Pending	Pending	Pending	
	VIT 19/2016	CATTLE	21-Sep-16	Pending	Pending	Pending	
	VIT 20/2016	CATTLE	23-Sep-16	Pending	Pending	Pending	
	VIT 21/2016	CATTLE	01-Nov-16	Pending	Pending	Pending	
	TOTAL :	101					

# Abbreviations used in table

FMD(V)	Foot-and-mouth disease (virus)
FMDV GD	Genome detected
FMDV NGD	Genome not detected (samples submitted in Trizol, only rRT-PCR carried out)
VI/ELISA	FMDV serotype identified following virus isolation in cell culture and antigen ELISA
rRT-PCR	Real-time reverse transcription polymerase chain reaction on epithelial suspension for FMD (or SVD) viral genome
NVD	No foot-and-mouth disease, swine vesicular disease or vesicular stomatitis virus detected
NT	Not tested



# 4.3. Antigenic Characterisation

Antigenic characterisation of FMD field isolates by matching with vaccine strains by 2dmVNT from October to December 2016.

Strain	Serotype	Topotype	Strain	O 3039	O1 Manisa	O/TUR/5/09
AFG/04/2016	0	ME-SA	PanAsia-2 <sup>ANT-10</sup>	М	М	М
AFG/12/2016	0	ME-SA	PanAsia-2 <sup>ANT-10</sup>	М	М	М
AFG/15/2016	0	ME-SA	PanAsia-2 <sup>ANT-10</sup>	М	М	М
AFG/16/2016	0	ME-SA	PanAsia-2 <sup>ANT-10</sup>	М	М	М
MAY/1/2015	0	SEA	Mya-98	М	М	М
MAY/10/2016	0	SEA	Mya-98	М	М	М
MAY/17/2014	0	SEA	Mya-98	М	М	М
MAY/5/2016	0	SEA	Mya-98	М	М	М
MUR/06/2016	0	ME-SA	Ind-2001d	М	М	М
MUR/07/2016	0	ME-SA	Ind-2001d	М	М	М
NIG/01/2016	0	unknown	unknown	N	N	В
NIG/04/2016	0	unknown	unknown	Ν	Ν	В
NIG/12/2016	0	unknown	unknown	М	М	М
NIG/19/2016	0	unknown	unknown	М	М	М
SAU/18/2016	0	ME-SA	PanAsia-2 <sup>ANT-10</sup>	М	М	М
TAI/26/2016	0	SEA	Mya-98	М	N	В
TAI/37/2016	0	SEA	Mya-98	М	N	М

Table 4: Vaccine matching studies for O FMDV by VNT

#### Table 5: Vaccine matching studies for A FMDV by VNT

Strain	Serotype	Topotype	Strain	A Iran-05	A/TUR/20/2006	A22 IRQ
AFG/5/2016	А	ASIA	Iran-05 <sup>FAR-11</sup>	Ν	Ν	N
MAY/15/2014	А	ASIA	Sea-97	N	Ν	М
SAU/19/2016	А	ASIA	G-VII	Ν	Ν	Ν
TAI/20/2016	А	ASIA	Sea-97	N	Ν	Ν
TAI/23/2016	А	ASIA	Sea-97	М	Ν	М

- A/MAY/15/2014 also matched against the vaccine virus A/MAY/97.
- A/SAU/19/2016 did not match against A/SAU/95
- A/AFG/5/2016 also matched against A/SAU/95



Strain	Serotype	Topotype	Strain	Asia 1 Shamir
AFG/6/2016	ASIA 1	ASIA	Sindh-08	М
AFG/10/2016	ASIA 1	ASIA	Sindh-08	М

#### Table 7: Vaccine matching studies for SAT 1 FMDV by VNT

Strain	Serotype	Topotype	Strain	SAT 1/RHO/12/78
NIG/1/2015	SAT 1	Х	-	N
NIG/2/2015	SAT 1	Х	-	Ν

# Abbreviations used in tables

М	Vaccine Match $r_1 = \ge 0.3$ . Suggests that there is a close relationship between field isolate and vaccine strain. A potent vaccine containing the vaccine strain is likely to confer protection.
Ν	No Vaccine Match $r_1 = < 0.3$ . Suggests that the field isolate is so different from the vaccine strain that the vaccine is unlikely to protect
В	Borderline Any r <sub>1</sub> values between 0.28 to 0.32
NT	Not tested against this vaccine



# 5. Annex 2

Recent FMD Publications (October to December 2016) cited by Web of Science (Pirbright Institute papers and authors are highlighted in **BOLD AND GREY**)

- 1. Abbishu, T., M. Araya, and R. Tesfaye (2016). Sero-prevalence status of foot and mouth disease in the North Western Amhara Regional State, Ethiopia. *Ethiopian Veterinary Journal*, **20**(2): 43-53.
- 2. Ali, M.R., H. Ullah, M.A. Siddique, M. Sultana, and M.A. Hossain (2016). Complete Genome Sequence of Pig-Originated Foot-and-Mouth Disease Virus Serotype O from Bangladesh. *Genome Announcements*, **4**(5).
- 3. Amaral, T.B., V. Gond, and A. Tran (2016). Mapping the likelihood of foot-andmouth disease introduction along the border between Brazil and Paraguay. *Pesquisa Agropecuaria Brasileira*, **51**(5): 661-670.
- 4. Asamenew, T., S. Mesfin, A. Ashebir, M. Ayelech, and G. Daniel (2016). Seroprevalence of foot and mouth disease in cattle in Borena Zone, Oromia regional state, Ethiopia. *Ethiopian Veterinary Journal*, **20**(1): 55-66.
- Bachanek-Bankowska, K., H.R. Mero, J. Wadsworth, V. Mioulet, R. Sallu, G.J. Belsham, C.J. Kasanga, N.J. Knowles, and D.P. King (2016). Development and evaluation of tailored specific real-time RT-PCR assays for detection of footand-mouth disease virus serotypes circulating in East Africa. *Journal of Virological Methods*, 237: 114-120.
- 6. Barkakati, J., S. Sarma, D.J. Kalita, J. Goswami, and K. Sharma (2016). Activity of different serum enzymes and hormone profile in foot and mouth disease affected cattle. *Indian Journal of Animal Research*, **50**(5): 826-827.
- Blanco, E., D. Andreu, and F. Sobrino, *Peptide vaccines against foot-and-mouth disease*. Foot-and-mouth disease virus: current research and emerging trends, ed. F. Sobrino and E. Domingo. 2017. 317-331.
- 8. Bouquet, B. (2016). Foot and mouth disease: pigs beware! *Point Veterinaire*, **47**(369 (Part 1)): 70-74.
- 9. Delgado, J., S. Pollard, K. Pearn, E.L. Snary, E. Black, G. Prpich, and P. Longhurst (2016). U.K. Foot and Mouth Disease: A Systemic Risk Assessment of Existing Controls. *Risk Analysis : an official publication of the Society for Risk Analysis*.
- Delirezh, N., R. Norian, and A. Azadmehr (2016). Changes in some pro-and antiinflammatory cytokines produced by bovine peripheral blood mononuclear cells following foot and mouth disease vaccination. *Archives of Razi Institute*, **71**(3): 199-208.
- Diaz-San Segundo, F., N.A. Montiel, D.F. Sturza, E. Perez-Martin, D. Hickman, E. Ramirez-Medina, M.J. Grubman, and T. de Los Santos (2016). Combination of Adt-O1Manisa and Ad5-bolFNλ3 induces early protective immunity against foot-and-mouth disease in cattle. *Virology*, **499**: 340-349.



- Domingo, E., I.d.I. Higuera, E. Moreno, A.I.d. Avila, R. Agudo, A. Arias, and C. Perales, *Quasispecies dynamics taught by natural and experimental evolution of foot-and-mouth disease virus*. Foot-and-Mouth Disease Virus: current research and emerging trends, ed. F. Sobrino and E. Domingo. 2017. 147-170.
- 13. Dong, Y.-m., J.-c. Cai, H.-t. Chen, and L. Chen (2016). Protection of a novel epitope-RNA VLP double-effective VLP vaccine for foot-and-mouth disease. *Antiviral Research*, **134**: 108-116.
- 14. Fernandez, P.J. and W.R. White, *Atlas of Transboundary Animal Diseases*. Atlas of Transboundary Animal Diseases, ed. P.J. Fernandez and W.R. White. 2016. 279 pp.-279 pp.
- 15. Fukai, K., T. Nishi, N. Shimada, K. Morioka, M. Yamada, K. Yoshida, K. Sakamoto, R. Kitano, R. Yamazoe, and M. Yamakawa (2016). Experimental infections using the foot-and-mouth disease virus O/JPN/2010 in animals administered a vaccine preserved for emergency use in Japan. *The Journal of Veterinary Medical Science*.
- Grant, C.F.J., B.V. Carr, N.B. Singanallur, J. Morris, S. Gubbins, P. Hudelet, M. Ilott, C. Charreyre, W. Vosloo, and B. Charleston (2016). The B-cell response to foot-and-mouth-disease virus in cattle following vaccination and live-virus challenge. *The Journal of General Virology*, 97(9): 2201-9.
- He, X., X. Lin, J. Liu, B. Wang, W. Zhuo, Y. Zhang, and Y. Zheng (2016). Adenoviral-mediated expression of type O foot-and-mouth disease viral polyprotein gene in mammary epithelial cells of diary goats. *Chinese Journal of Veterinary Science*, **36**(9): 1537-1543.
- Hernandez-Jover, M., N. Schembri, P.K. Holyoake, J.-A.L.M.L. Toribio, and P.A.J. Martin (2016). A Comparative Assessment of the Risks of Introduction and Spread of Foot-and-Mouth Disease among Different Pig Sectors in Australia. *Frontiers in Veterinary Science*, **3**: 85-85.
- Hiesel, J.A., I. Kopacka, R. Fuchs, H. Schobesberger, P. Wagner, A. Loitsch, and J. Kofer (2016). Epidemiological evaluation of different FMD control strategies in two selected regions in Austria. *Berliner und Munchener Tierarztliche Wochenschrift*, **129**(11/12): 484-494.
- Huang, Y., H. Zang, Y. Yu, Q. Wang, Z. Fu, and leee. Soft-Sensor Modeling of Foot-and-Mouth Disease Vaccine Suspension Culture Based on Relevance Vector Machine, in Proceedings of the 28<sup>th</sup> Chinese Control and Decision Conference. 2016. p. 5524-5529.
- 21. Jemberu, W.T., M. Mourits, J. Rushton, and H. Hogeveen (2016). Cost-benefit analysis of foot and mouth disease control in Ethiopia. *Preventive Veterinary Medicine*, **132**: 67-82.
- Jemberu, W.T., M.C.M. Mourits, M. Sahle, B. Siraw, J.C.M. Vernooij, and H. Hogeveen (2016). Epidemiology of Foot and Mouth Disease in Ethiopia: a Retrospective Analysis of District Level Outbreaks, 2007-2012. *Transboundary and Emerging Diseases*, 63(6): e246-e259.



- 23. Kardjadj, M. (2016). Foot-and-mouth disease (FMD) in the Maghreb and its threat to southern European countries. *Tropical Animal Health and Production*.
- 24. Kardjadj, M. and P.D. Luka (2016). Molecular epidemiology of foot and mouth disease, bluetongue and pest de petites ruminants in Algeria: historical perspective, diagnosis and control. *African Journal of Biotechnology*, **15**(44): 2474-2479.
- 25. Lawrence, P. and E. Rieder, *Foot-and-mouth disease virus receptors: multiple gateways to initiate infection.* Foot-and-mouth disease virus: current research and emerging trends, ed. F. Sobrino and E. Domingo. 2017. 107-135.
- 26. Loundras, E.-A., M.R. Herod, M. Harris, and N.J. Stonehouse (2016). Foot-andmouth disease virus genome replication is unaffected by inhibition of type III phosphatidylinositol-4-kinases. *Journal of General Virology*, **97**: 2221-2230.
- Loundras, E.-A., M.R. Herod, M. Harris, and N.J. Stonehouse (2016). Foot-andmouth disease virus genome replication is unaffected by inhibition of type III phosphatidylinositol-4-kinases. *The Journal of General Virology*, **97**(9): 2221-30.
- Ludi, A., V. Mioulet, N.J. Knowles, and D.P. King, Laboratory diagnostic methods to support the surveillance and control of foot-and-mouth disease. Footand-Mouth Disease Virus: current research and emerging trends, ed. F. Sobrino and E. Domingo. 2017. 275-285.
- 29. Ma, X.X., Y.P. Feng, Y.X. Gu, J.H. Zhou, and Z.R. Ma (2016). Effect of the nucleotides surrounding the start codon on the translation of foot-and-mouth disease virus RNA. *Acta Virologica*, **60**(2): 151-155.
- Madhanmohan, M., S. Yuvaraj, K. Manikumar, R. Kumar, S.B. Nagendrakumar, S.K. Rana, and V.A. Srinivasan (2016). Evaluation of the Flinders Technology Associates Cards for Storage and Temperature Challenges in Field Conditions for Foot-and-Mouth Disease Virus Surveillance. *Transboundary and Emerging Diseases*, **63**(6): 675-680.
- 31. Mahy, B.W.J. and G.J. Belsham, *Overview of foot-and-mouth disease and its impact as a re-emergent viral infection*. Foot-and-Mouth Disease virus: current research and emerging trends, ed. F. Sobrino and E. Domingo. 2017. 417-426.
- Martinez-Salas, E. and G.J. Belsham, Genome organization, translation and replication of foot-and-mouth disease virus RNA. Foot-and-Mouth Disease virus: current research and emerging trends, ed. F. Sobrino and E. Domingo. 2017. 13-42.
- 33. Mateu, M.G., *The foot-and-mouth disease virion: structure and function*. Footand-Mouth Disease Virus: current research and emerging trends, ed. F. Sobrino and E. Domingo. 2017. 61-105.
- McCullough, K.C., M. Saiz, and A. Summerfield, *Innate to adaptive: immune defence handling of foot-and-mouth disease virus*. Foot-and-Mouth Disease virus: current research and emerging trends, ed. F. Sobrino and E. Domingo. 2017. 211-274.



- 35. Nfon, C., O. Lung, C. Embury-Hyatt, and S. Alexandersen, *Clinical signs and pathology of foot-and-mouth disease*. Foot-and-Mouth Disease virus: current research and emerging trends, ed. F. Sobrino and E. Domingo. 2017. 171-178.
- 36. Pacheco, J.M., C. Stenfeldt, L.L. Rodriguez, and J. Arzt (2016). Infection Dynamics of Foot-and-Mouth Disease Virus in Cattle Following Intranasopharyngeal Inoculation or Contact Exposure. *Journal of Comparative Pathology*.
- 37. Park, H., S. Bae, and 박선일 (2016). Properties of a Social Network Topology of Livestock Movements to Slaughterhouse in Korea. *Journal of Veterinary Clinics*, **33**(5): 278-285.
- Park, J.-N., M.-K. Ko, R.-H. Kim, M.-E. Park, S.-Y. Lee, J.-E. Yoon, J.-H. Choi, S.-H. You, J.-W. Park, K.-N. Lee, J.-E. Chun, S.-M. Kim, D. Tark, H.-S. Lee, Y.-J. Ko, B. Kim, M.-H. Lee, and J.-H. Park (2016). Construction of stabilized and tagged foot-and-mouth disease virus. *Journal of Virological Methods*, 237: 187-191.
- Patterson, G.R., A.H. Mohr, T.P. Snider, T.A. Lindsay, P.R. Davies, T.J. Goldsmith, and F. Sampedro (2016). Prioritization of Managed Pork Supply Movements during a FMD Outbreak in the US. *Frontiers in Veterinary Science*, **3**: 97-97.
- Pillay, P., K.J. Kunert, S. van Wyk, M.E. Makgopa, C.A. Cullis, and B.J. Vorster (2016). Agroinfiltration contributes to VP1 recombinant protein degradation. *Bioengineered*, 7(6): 459-477.
- Pozzi, P., M. Etinger, B. Gelman, V. Pirogov, E. Khinich, and Y. Hadani. Clinical description of an outbreak of foot and mouth disease in a close-cycle unit, in Atti della Societa Italiana di Patologia ed Allevamento dei Suini, XLII Meeting Annuale, Montichiari, Italia, 10-11 Marzo 2016. 2016. p. 139-145.
- Rhyan, J., M. McCollum, T. Gidlewski, M. Shalev, G. Ward, B. Donahue, J. Arzt, C. Stenfeldt, F. Mohamed, P. Nol, M. Deng, S. Metwally, T. McKenna, and M. Salman (2016). Foot-and-Mouth Disease in a small sample of experimentally infected pronghorn (*Antilocapra americana*). *Journal of Wildlife Diseases*, **52**(4): 862-873.
- 43. Rodenbusch, C.R., L.R. da Silveira, A.R. Bavaresco, and M.V. Burgel Sfoggia (2016). An Assessment of Microbiological Methods to Test Sterility of Foot-and-mouth Disease Vaccines Produced in Brazil. *Acta Scientiae Veterinariae*, **44**.
- 44. Rowlands, D.J., *Introduction: foot-and-mouth disease much progress but still a lot to learn*. Foot-and-Mouth Disease Virus: current research and emerging trends, ed. F. Sobrino and E. Domingo. 2017. 1-12.
- 45. Sanson, R.L., T. Rawdon, K. Owen, K. Hickey, M. van Andel, and Z.D. Yu (2016). Evaluating the benefits of vaccination when used in combination with stampingout measures against hypothetical introductions of foot-and-mouth disease into New Zealand: a simulation study. *New Zealand Veterinary Journal*: 1-28.



- Segundo, F.D.-S., N.A. Montiel, D.F. Sturza, E. Perez-Martin, D. Hickman, E. Ramirez-Medina, M.J. Grubman, and T. de los Santosa (2016). Combination of Adt-O1Manisa and Ad5-bolFN lambda 3 induces early protective immunity against foot-and-mouth disease in cattle. *Virology*, **499**: 340-349.
- 47. Shakiba, Y., S.E. Rezatofighi, S.M.S. Nejad, and M.R. Ardakani (2016). Antiviral activity of Alhagi maurorum Medik's methanolic extract on foot and mouth disease virus (FMDV) in cell cultures. *Jundishapur Journal of Natural Pharmaceutical Products*, **11**(3): e30641-e30641.
- 48. Shi, Q., J. Feng, Y. Chen, X. Bai, T. Cui, J. Cui, L. Wang, and G. Zhang (2016). Sequence analysis 5 strains of VP1 and 3A genes from foot-and-mouth disease virus. *Zhongguo Yufang Shouyi Xuebao*, **38**(8): 609-613.
- 49. Shu, J., S. Li, H. Shen, and Y. Guo (2016). Screening of fusion tags that enhanced expression and solubility of type A foot-and-mouth disease virus VP1 protein. *Journal of Henan Agricultural Sciences*, **45**(9): 114-119.
- 50. Slack (2016). Pharmacological characterization of the av beta 6 integrin binding and internalization kinetics of the foot-and-mouth disease virus derived peptide A20FMDV2 (vol 97, pg 114, 2016). *Pharmacology*, **98**(5-6): 228-228.
- Smitsaart, E.N. and I.E. Bergmann, Quality attributes of current inactivated footand-mouth disease vaccines and their effects on the success of vaccination programmes. Foot-and-Mouth Disease Virus: current research and emerging trends, ed. F. Sobrino and E. Domingo. 2017. 287-316.
- 52. Sobrino, F. and E. Domingo, *Foot-and-Mouth Disease Virus: current research and emerging trends*. Foot-and-Mouth Disease Virus: current research and emerging trends, ed. F. Sobrino and E. Domingo. 2017. xxii + 431 pp.-xxii + 431 pp.
- Sun, Y., M. Ma, G. Zhou, M. Lin, Q. Zeng, and T. Jiang (2016). Development of gold-labeled immunochromatographic test strip for detecting antibodies against serotype A foot-and-mouth disease virus. *Chinese Veterinary Science*, **46**(9): 1123-1127.
- 54. Thwiny, H.T. (2016). Diagnosis of carrier state of foot and mouth disease virus in vaccinated and unvaccinated cattle by RT-PCR. *Basrah Journal of Veterinary Research*, **15**(2): 30-38.
- Tildesley, M.J., W.J.M. Probert, and M.E.J. Woolhouse, *Mathematical models of the epidemiology and control of foot-and-mouth disease*. Foot-and-Mouth Disease Virus: current research and emerging trends, ed. F. Sobrino and E. Domingo. 2017. 385-408.
- 56. Tulloch, F., G.A. Luke, and M.D. Ryan, *Foot-and-mouth disease virus proteinases and polyprotein processing*. Foot-and-Mouth Disease Virus: current research and emerging trends, ed. F. Sobrino and E. Domingo. 2017. 43-59.
- Ularamu, H.G., J.O. Ibu, B.A. Wood, J.N. Abenga, D.D. Lazarus, Y.S. Wungak, N.J. Knowles, J. Wadsworth, V. Mioulet, D.P. King, D. Shamaki, and M.I. Adah (2016). Characterization of Foot-and-Mouth Disease Viruses Collected in Nigeria



Between 2007 and 2014: Evidence for Epidemiological Links Between West and East Africa. *Transboundary and Emerging Diseases*.

- 58. Vallat, B., J. Domenech, and A.A. Schudel, *The role of international organizations in the control of foot-and-mouth disease*. Foot-and-Mouth Disease Virus: current research and emerging trends, ed. F. Sobrino and E. Domingo. 2017. 409-416.
- 59. Van Phan, L., V. Thi Thu Hang, D. Hong-Quan, T. Van Thai, and D. Song (2016). Evolutionary phylodynamics of foot-and-mouth disease virus serotypes O and A circulating in Vietnam. *BMC Veterinary Research*, **12**.
- 60. Vleeschauwer, A.R.d., D.J. Lefebvre, and K.d. Clercq, *Antiviral therapies for footand-mouth disease*. Foot-and-Mouth Disease Virus: current research and emerging trends, ed. F. Sobrino and E. Domingo. 2017. 357-383.
- 61. Vosloo, W. and G.R. Thomson, *Natural habitats in which foot-and-mouth disease viruses are maintained*. Foot-and-Mouth Disease Virus: current research and emerging trends, ed. F. Sobrino and E. Domingo. 2017. 179-210.
- 62. Wang, H.M., X.Z. Xia, G.X. Hu, L. Yu, and H.B. He (2016). Bovine Mx1 enables resistance against foot-and-mouth disease virus in naturally susceptible cells by inhibiting the replication of viral RNA. *Acta Virologica*, **60**(1): 85-93.
- 63. Yang, M., W. Xu, H. Bittner, J. Horsington, W. Vosloo, M. Goolia, D. Lusansky, and C. Nfon (2016). Generation of mAbs to foot-and-mouth disease virus serotype A and application in a competitive ELISA for serodiagnosis. *Virology Journal*, **13**.



# 6. Annex 3

RECOMMENDATIONS FROM WRLFMD® ON FMD VIRUS STRAINS TO BE INCLUDED IN FMDV ANTIGEN BANKS (FOR FMD-FREE COUNTRIES) December 2016

Note: Virus strains are NOT listed in order of importance

	A/ASIA/G-VII(G-18)*				
	· · · · ·				
	O Manisa				
Lliada	O PanAsia-2 (or equivalent)				
High	O BFS or Campos				
Priority	A24 Cruzeiro				
· · · · · · · · · · · · · · · · · · ·	Asia 1 Shamir				
	A Iran-05 <i>(or A TUR 06)</i>				
	A22 Iraq				
	SAT 2 Saudi Arabia (or equivalent i.e. SAT 2 Eritrea)				
	A Eritrea				
	SAT 2 Zimbabwe				
Medium	SAT 1 South Africa				
	A Malaysia 97 (or Thai equivalent such as A/Sakolnakorn/97)				
Priority	A Argentina 2001				
	O Taiwan 97 (pig-adapted strain or Philippine				
	equivalent)				
	A Iran '96				
	A Iran '99				
	A Iran 87 or A Saudi Arabia 23/86 (or equivalent)				
Low	A15 Bangkok related strain				
	A87 Argentina related strain				
Priority	C Noville				
	SAT 2 Kenya				
	5				
	SAT 3 Zimbabwe				
	SAT 1 Kenya				

Note: Discussions are currently underway to adopt a risk-based approach for different FMD viral lineages to identify priority vaccines for use in Europe and other FMD-free settings.

\*Recent *in vitro* data from WRLFMD for serotype A viruses from Saudi Arabia and Iran highlights an apparent gap in vaccine coverage. Work is urgently required to evaluate whether there is adequate *in vitro* match with Indian vaccine strains (A/IND/40/2000), or whether *in vivo* protection may be provided by high potency international vaccines.